EXPRESSION OF GLYPICAN-3(GPC-3) AND 8-HYDROXYDEOXYGUANOSINE (8-OHDG) IN CHRONIC HEPATITIS C AND RELATIONSHIP WITH HEPATOCELLULAR CARCINOMA

El-Alfy Y, El-Kashishy K, George M, Abdel-Wahab M
*Pathology Department, Faculty of Medicine, Zagazig University

ABSTRACT

Background: In Egypt, hepatocellular carcinoma (HCC) is the second most common cancer in men and the 6th most common cancer in women [1]. Egypt has the highest prevalence of HCV in the world and the prevalence of HCC is increasing in the last years [2]. Nowadays, HCV infection and its complications are among the leading public health challenges in Egypt with 13.8% of population infected [3] and in these patients, the risk of HCC is increased 17-fold [4]. Glypican-3 (GPC-3) is a member of heparin sulfate proteoglycan family which is linked to cell surface. It plays an important role in regulation of cell growth, differentiation, and migration. It is normally expressed in fetal liver and placenta but not in normal adult liver tissue [5].

Data have demonstrated a high incidence of GPC-3 expression in HCC and suggested its implication in detecting malignant hepatic lesions and it is widely recognized as an efficient serological and histochemoical marker for early hepatocellular carcinoma [6,7]. However, GPC-3 expression can also be detected in a variable percentage of benign liver tissue with active hepatitis, regenerative nodules and high-grade dysplastic nodules [6,8,9].

8-Hydroxydeoxyguanosine (8-OHdG) is an oxidatively modified promutagenic DNA that is produced by oxygen radicals and is recognized as a useful marker in estimating DNA damage induced by oxidative stress [10]. 8-Hydroxydeoxyguanosine is one of the most widely used oxidative stress biomarkers, mainly because of its abundance in DNA and also because of its reliable detectability [11]. In patients with chronic hepatitis C, increased 8-OHdG in DNA extracted from liver tissue was reported [12]. Studies have shown that high expression of 8-OHdG in livers with chronic hepatitis C predicted the development of primary HCC [13,14] as free radical production is increased at the site of inflammation, resulting in lipid peroxidation and oxidative DNA damage, which are risk factors for HCC suggesting the association of 8-OHdG and carcinogenesis in livers with chronic hepatitis C [15].

INTRODUCTION

In Egypt, hepatocellular carcinoma (HCC) is the second most common cancer in men and the 6th most common cancer in women [1]. Egypt has the highest prevalence of HCV in the world and the prevalence of HCC is increasing in the last years [2].

Nowadays, HCV infection and its complications are among the leading public health challenges in Egypt with 13.8% of population infected [3] and in these patients, the risk of HCC is increased 17-fold [4].

Glypican-3 (GPC-3) is a member of heparin sulfate proteoglycan family which is linked to cell surface. It plays an important role in regulation of cell growth, differentiation, and migration. It is normally expressed in fetal liver and placenta but not in normal adult liver tissue [5].

Data have demonstrated a high incidence of GPC-3 expression in HCC and suggested its implication in detecting malignant hepatic lesions and it is widely recognized as an efficient serological and histochemoical marker for early hepatocellular carcinoma [6,7].
MATERIAL AND METHODS

This work is a retrospective, cross-sectional study carried out on 75 liver paraffin blocks that were previously diagnosed as chronic hepatitis C (50 cases) and hepatocellular carcinoma (25 cases), collected from the Pathology Department, Faculty of Medicine, Zagazig University, National Cancer Institute in Egypt and liver cancer institute in Menofeya, in the period from October 2012 to January 2014. The selected specimens were obtained by needle biopsy and surgical excision.

The clinical data concerning age and sex were obtained from the patients files.

Paraffin blocks of all cases were sectioned at 3-5 micron thickness and stained with routine hematoxylin and eosin stain to re-evaluate and confirm the diagnosis.

A) The histological criteria of HCV were assessed according to METAVIR system [16].

B) HCC cases were graded into well, moderately and poorly differentiated according to WHO [17].

The inclusion and exclusion criteria used in the current study were:
1) All cases of hepatocellular carcinoma were on top of chronic hepatitis C and cirrhosis.
2) Pathological diagnosis of hepatocellular lesions consistent with histological diagnostic criteria of WHO.
3) Chronic hepatitis B, alcoholic hepatitis and autoimmune hepatitis were excluded.
4) Fragmented core biopsies or core biopsies containing less than 6 portal tracts were excluded.

Histopathologic examination:

The 50 cases of chronic hepatitis C included 29 cases low grade chronic hepatitis (A1) and 21 cases high grade chronic hepatitis (9 cases A2 and 12 cases A3). 15 cases were cirrhotic.

The 25 HCC cases included 9 cases well differentiated (Grade I), 11 moderately differentiated (Grade II) and 5 poorly differentiated (Grade III). Among them, 22 cases showed cirrhosis in the non neoplastic liver tissue.

Immunochemical staining:

Immunohistochemical staining was carried out using the streptavidin–biotin immunoperoxidase technique (Dako-Cytomation, Glostrup, Denmark). Sections of 3–5 mm thickness were cut from formalin-fixed, paraffin-embedded blocks, mounted on positively charged slides, deparaffinized in xylene, and rehydrated in graded alcohol. Sections were boiled in citrate buffer (pH 6.0) for 20 minutes and then washed in PBS (pH 7.3). Thereafter, blocking of endogenous peroxidase activity with 6% H2O2 in methanol was carried out. The slides were then incubated overnight with monoclonal antibodies: Glypican-3; Mouse monoclonal antibody, Dilution 1:100; (CM396, A.B, Clone IG12, Biocare Medical LLC, Concord, USA); 8-Hydroxydeoxyguanosine(8-OHdG); Monoclonal antibody, Dilution 1:100 (Clone15A3, SC-66036, Santa Cruz, California, USA). Incubation with a secondary antibody and product visualization were performed (Dako-Cytomation) with diaminobenzidine substrate (Research Genetics, Huntsville, Alabama, USA) as the chromogen. The slides were finally counterstained with Mayer’s hematoxylin (BioGenex Laboratories, San Ramon, California, USA) and washed once each with distilled water and PBS. Positive and negative controls were stained at the same staining setting with the studied cases: Hepatocellular carcinoma and Chronic hepatitis C were used as positive control for glypican-3 and 8-OHdG respectively, while negative controls were done using the same tissue with omission of the primary antibody.

Evaluation of the results of immunohistochemical staining:

1- Glypican-3 immunostaining:

The GPC-3 staining was considered positive when the granular brown reaction was found in the cytoplasm and/or the membranes. Immunoreactivity was determined semiquantitatively by examining fields (magnification, x200). Using a (0–3+) scale, the staining was described as 0 staining (negative), 1+ staining (<10% of cells), 2+staining (10%–25% of cells), or 3+staining (>25% of cells)[18].

2- 8-Hydroxydeoxyguanosine:

The number of 8-OHdG stained hepatocytes (nuclear stain) in 10 random fields (magnification, x400) was counted. The percentage of positively stained cells was calculated, which was termed the 8-OHdG label index (LI), in each field and graded as follow: low, LI <50%; high, LI >50% [15].

The slides were evaluated by three different pathologists working separately. All discrepancies were discussed and consensus reached.

Statistical analysis:

Statistical analysis was performed using SPSS software (SPSS, Chicago, IL, USA). Data were expressed as mean ±SD for quantitative variables. For categorical variables Fisher’s exact test or chi-square was used. P-value less than 0.05 was considered significant.

RESULTS

The age of the studied cases of chronic hepatitis C ranged from (28-62) and the mean was 40.3± 6.8 year. The majority of studied cases of chronic hepatitis C were below 39 years (48%).
The age of the studied cases of HCC ranged from (41-73) and the mean was 57.4±8.1. The majority of studied cases of HCC were between 50-59 (44%). Glypican-3 expression was observed in 18 % of cases of chronic hepatitis C, all positive cases were in the high grade (42.9%), while none of low grade chronic hepatitis cases showed glypican-3 positivity. There was a statistically significant difference in glypican-3 expression between low and high grade chronic hepatitis C (P=0.0016) (Table 1, Fig. 1).

Glypican-3 expression was observed in 88% of HCC cases (77.7% of well differentiated HCC, 100% of moderately differentiated HCC and in 80% of poorly differentiated HCC). There was no statistically significant difference in glypican-3 expression among different grades (Table 2, Fig. 2).

High 8-OHdG index was detected in 72.4% of cases of low grade chronic hepatitis and 95.2% of cases of high grade chronic hepatitis, while low index was detected in 27.6% of low grade chronic hepatitis and in 4.8% of cases of high grade chronic hepatitis. There was a statistically significant difference in 8-OHdG expression between low and high grade chronic hepatitis C (P=0.05) (Table 3, Fig. 3).

High 8-OHdG index was observed in 52% of cases of HCC while low index was observed in 48% of cases. There was no statistically significant difference in 8-OHdG expression between different grades of HCC (P= 0.83) (Table 4, Fig. 4).

**Table (1): Immunoreactivity pattern of GPC-3 in chronic hepatitis C according to grade:**

<table>
<thead>
<tr>
<th>Chronic hepatitis C</th>
<th>GPC-3 immunoreactivity</th>
<th>Total Positive</th>
<th>$X^2$</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Negative (0)</td>
<td>Positive (+)</td>
<td>Positive (+++)</td>
<td>Positive (+++)</td>
</tr>
<tr>
<td>Low grade hepatitis</td>
<td>No</td>
<td>%</td>
<td>No</td>
<td>%</td>
</tr>
<tr>
<td>29</td>
<td>100</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>21</td>
<td>57.1</td>
<td>23.8</td>
<td>9.5</td>
<td>2</td>
</tr>
<tr>
<td>50</td>
<td>82.0</td>
<td>10.0</td>
<td>2</td>
<td>4.0</td>
</tr>
</tbody>
</table>

* Significant (P < 0.05).

**Table (2): Immunoreactivity pattern of GPC-3 in hepatocellular carcinoma:**

<table>
<thead>
<tr>
<th>Hepatocellular carcinoma</th>
<th>GPC-3 immunoreactivity</th>
<th>Total Positive</th>
<th>$X^2$</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Negative expression (0)</td>
<td>Positive expression (+)</td>
<td>Positive expression (+++)</td>
<td>Positive expression (+++)</td>
</tr>
<tr>
<td>Well differentiated HCC (Grade I)</td>
<td>No</td>
<td>%</td>
<td>No</td>
<td>%</td>
</tr>
<tr>
<td>9</td>
<td>22.2</td>
<td>44.4</td>
<td>2</td>
<td>22.2</td>
</tr>
<tr>
<td>Moderately differentiated HCC (Grade II)</td>
<td>HCC</td>
<td>0</td>
<td>9.0</td>
<td>5</td>
</tr>
<tr>
<td>Poorly differentiated HCC (Grade III)</td>
<td>5</td>
<td>12.0</td>
<td>24.0</td>
<td>8</td>
</tr>
<tr>
<td>Total</td>
<td>25</td>
<td>12.0</td>
<td>6</td>
<td>24.0</td>
</tr>
</tbody>
</table>

**Table (3): Immunoreactivity pattern of 8-OHdG in chronic hepatitis C:**

<table>
<thead>
<tr>
<th>Chronic hepatitis C</th>
<th>Low index of 8-OHdG expression</th>
<th>High index of 8-OHdG expression</th>
<th>$X^2$</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>No</td>
<td>%</td>
<td>No</td>
<td>%</td>
<td>No</td>
</tr>
<tr>
<td>Low grade hepatitis</td>
<td>29</td>
<td>8</td>
<td>27.6</td>
<td>21</td>
</tr>
<tr>
<td>High grade hepatitis</td>
<td>21</td>
<td>1</td>
<td>4.8</td>
<td>20</td>
</tr>
<tr>
<td>Total</td>
<td>50</td>
<td>9</td>
<td>18</td>
<td>41</td>
</tr>
</tbody>
</table>

* Significant (P = 0.05).
**Table (4): Immunoreactivity pattern of 8-OHdG in heptocellular carcinoma:**

<table>
<thead>
<tr>
<th>Heptocellular carcinoma</th>
<th>Low index of 8-OHdG expression</th>
<th>High index of 8-OHdG expression</th>
<th>( \chi^2 )</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>No</td>
<td>%</td>
<td>No</td>
<td>%</td>
</tr>
<tr>
<td>Well differentiated HCC (Grade I)</td>
<td>9</td>
<td>55.6</td>
<td>4</td>
<td>44.5</td>
</tr>
<tr>
<td>Moderately differentiated HCC (Grade II)</td>
<td>11</td>
<td>44.5</td>
<td>6</td>
<td>54.5</td>
</tr>
<tr>
<td>Poorly differentiated HCC (Grade III)</td>
<td>5</td>
<td>40</td>
<td>3</td>
<td>60</td>
</tr>
<tr>
<td>Total</td>
<td>25</td>
<td>48%</td>
<td>13</td>
<td>52%</td>
</tr>
</tbody>
</table>

\[ X^2 = 0.36 \quad P = 0.83 \]

**Fig. 1: Glypican-3 immunoreactivity in chronic hepatitis C:**

- A core needle biopsy of chronic hepatitis C (A2, F3) showing positive glypican-3 cytoplasmic immunoreactivity (Score 3+) (Immunoperoxidase stain, X 200).
- Higher magnification of previous image showing positive glypican-3 cytoplasmic immunoreactivity (Score 3+) (Immunoperoxidase stain, X 400).
- A core needle biopsy of chronic hepatitis C (A3, F4) showing positive glypican-3 cytoplasmic immunoreactivity (Score 3+) (Immunoperoxidase stain, X 100).
Fig. 2: Glypican-3 immunoreactivity in hepatocellular carcinoma:

a- Well differentiated HCC showing positive glypican-3 cytoplasmic immunoreactivity (score 3+)(Immunoperoxidase stain, X 400 ).

b- Moderately differentiated HCC (Clear cell type) showing positive glypican-3 membranous immunoreactivity (score 3+) (Immunoperoxidase stain, X 200 ).

c- Poorly differentiated HCC showing positive glypican-3 cytoplasmic immunoreactivity (score 3+)(Immunoperoxidase stain, X 300 ).
Fig. 3: 8-OhdG immunoreactivity in chronic hepatitis C:
A- A core needle biopsy of chronic hepatitis C (A2, F2) showing high index 8-OhdG nuclear immunoreactivity (Immunoperoxidase stain, X 200).
B- A core needle biopsy of chronic hepatitis C (A2, F3) showing high index 8-OhdG nuclear immunoreactivity (Immunoperoxidase stain, X 200).
C- A core needle biopsy of chronic hepatitis C (A3, F4) showing high index 8-OhdG nuclear immunoreactivity, also immunoreactivity of bile duct epithelium and inflammatory cells (Immunoperoxidase stain, X 300).
Fig. 4: 8-OHdG immunoreactivity in hepatocellular carcinoma:

a- Moderately differentiated HCC (acinar pattern) showing high index 8-OHdG nuclear immunoreactivity (Immunoperoxidase stain, X 400).

b- Moderately differentiated HCC (clear cell type) showing high index 8-OHdG nuclear immunoreactivity (Immunoperoxidase stain, X 300).

c- Poorly differentiated HCC showing high index 8-OHdG nuclear immunoreactivity (Immunoperoxidase stain, X 300).
DISCUSSION

The present work is a retrospective, cross-sectional study performed on 75 liver paraffin blocks that included 50 cases of chronic hepatitis C and 25 cases of hepatocellular carcinoma. These cases were collected from the Pathology Department, Faculty of Medicine, Zagazig University, National Cancer Institute in Egypt and liver cancer institute in Menofeya, in the period from October 2012 to January 2014.

The 50 cases of chronic hepatitis C included 29 cases low grade chronic hepatitis and 21 cases high grade chronic hepatitis, 15 patients had cirrhosis, all in the high-grade group. The 25 cases of hepatocellular carcinoma (HCC) included 9 cases well differentiated HCC (grade I), 11 cases moderately differentiated HCC (grade II) and 5 cases poorly differentiated HCC (grade III). Among 25 cases of HCC, 22 cases showed cirrhosis in the non neoplastic liver tissue.

Glypican-3 (GPC-3) belongs to glypican family that is a group of heparin sulfate proteoglycans linked to the outer surface of cell membrane through a glycosyl-phosphatidylinositol anchor [19].

In our study, Glypican-3 was expressed in 9 (18%) of 50 cases of chronic hepatitis, all of high grade hepatitis (42.9%) and non of low grade chronic hepatitis. The staining pattern was exclusively cytoplasmic without staining of plasma membranes or canaliculi, as has been described in some HCCs. There was a statistically significant difference between GPC-3 expression in low and high grade hepatitis (P=0.0016). This was consistent with Abdul-Al et al. [9] who studied GPC-3 expression in 60 cases of chronic hepatitis C (30 low grade and 30 high grade) and found that 25 (83.3%) of 30 cases of high grade chronic hepatitis showed positive staining for GPC-3. In contrast, all biopsies in the low-grade group were negative. The positive hepatocytes were present in small groups, usually haphazardly distributed but sometimes with a periportal predominance.

In the present study, glypican-3 was expressed in 13 (35.1%) of 37 cases of cirrhosis, 8/22 (36.4%) of cirrhosis with carcinoma and 5/15 (33.3%) of cirrhosis without carcinoma. On the other hand, Baumhoer et al. [8] found positivity in 11/95 (12%) of liver cirrhosis and Shafizadeh et al. [20] found focal positivity in 4 cases of 35 cases of cirrhosis (11%).

In this work, there was no statistical significant difference in glypican-3 expression between cirrhosis with or without carcinoma (P=0.97). However, Wang et al. [21] reported that among 67 HCCs with a cirrhotic background, focal GPC-3 immunoreactivity was detected in a small proportion of the cirrhotic nodules in 11 (16.4%) cases but not in cirrhotic livers without HCC.

Several studies have demonstrated that GPC-3 is positive in most HCC, and positive immunostaining has been detected in 52.5% to 100% of HCC [5,6, 20, 22, 23]. In the present study, glypican-3 was detected in 22 (88%) of 25 cases of HCC (77.7% (7/9) of well differentiated cases, 100% (11/11) of moderately differentiated cases, and 80% (4/5) of poorly differentiated cases) and there was no statistically significant difference in glypican-3 expression between different grades of HCC. Reactivation of the fetal phenotype, which is common in malignant tumors, may explain the expression of GPC3 in malignant hepatocellular nodules [24].

8-OHdG is considered a useful indicator for investigating the involvement of oxidative stress in hepatocarcinogenesis [25]. The expression of 8-OHdG have been reported in livers with chronic hepatitis C [26], cirrhosis [27] and carcinoma [28].

In our study, 8-OHdG was observed in all cases of chronic hepatitis C, with high labeling index in 41/50 (82%) of cases and low index in 9/50 (18%) of cases. The staining was seen mainly in nuclei of hepatocytes, bile duct cells, portal inflammatory cells, and occasionally in the nuclei of sinusoidal cells.

Similar results were reported by Mahmood et al. [29] who studied the expression of 8-OHdG in eight patients with chronic hepatitis C and found that 8-OHdG was observed in the nuclei of hepatocytes in all cases and Fujita et al. [30] who found immunoreactivity for 8-OHdG in all 77 (100%) of chronic hepatitis C examined patients.

On the other hand, Sumiyoshi et al. [31] observed positive staining for 8-OHdG in 64% of HCV patients, Chuma et al. [13] detected expression of 8-OHdG in 62 (59.6%) of 104 patients with chronic hepatitis and Kitada et al. [32] detected 8-OHdG expression in 17 of 24 cases (70.8%) with chronic hepatitis C.

In the present study, liver 8-OHdG expression was correlated with hepatic inflammation and there was a statistically significant difference in 8-OHdG expression between low and high grade hepatitis (P=0.05), this was consistent with Fujita et al. [33] who found that 8-OHdG expression was correlated with the grade of hepatitis (P = 0.032), this correlation may indicate the direct involvement of hepatic oxidative stress in the pathogenesis and...
progression of liver cell injury in chronic viral hepatitis.

This correlation with the grade of hepatitis was also consistent with the results of Chuma et al.[13] and Kitada et al.[32] who reported that 8-OHdG was correlated with the grade of hepatitis but in contrast to Fukushima et al.[34] who found that the number of 8-OHdG-positive cells was not correlated with hepatic inflammatory activity grade.

Our study showed high labeling index of 8-OHdG in 13/25 (52%) of cases of HCC and low index in 12/25 (68%) of cases and there was no statistically significant difference between grades, this positivity for 8-OHdG in HCC was also reported by Ichiba et al.[28] and Jo et al.[35] who observed 8-OHdG positivity in 25 (83%) of the 30 HCC cases and 56.4% of their examined HCC cases respectively.

In this work, there was a statistically significant difference in 8-OHdG expression between HCC and chronic hepatitis C (P=0.006), this was consistent with Fukushima et al., [34] who compared the expression levels of 8-OHdG in non-tumor liver tissue of HCC patients without diagnosed disease predisposing to those of HCC patients with chronic hepatitis C and found a significant higher expression level in those with HCV.

Our results also similar to Schwartz et al.[25] who demonstrated increased 8-OHdG concentrations in non tumorous tissue of HCC patients compared to tumorous tissue.

Previous reports have noted the high expression of 8-OHdG in the livers of chronic viral hepatitis patients [28,32], and our results are consistent with these reports. Recent studies have shown that high expression of 8-OHdG in livers with CHV predicted the development of primary HCC [13,14] and also postoperative recurrence [36,37], suggesting the association of 8-OHdG and carcinogenesis in livers with chronic viral hepatitis C. Our results demonstrated that oxidative DNA damage widely occurs in the liver with chronic liver injury suggesting a possible link between active inflammation and hepatocarcinogenesis.

The presence of oxidative stress markers in HCC tissue as well as in non-HCC tissue indicates that oxidative stress is closely related to the development of HCC and that non-HCC tissue under oxidative stress may become carcinomatous in the future. The importance of detection of OS markers and antioxidant therapy in HCV-associated liver disease is to slow down disease progression and HCC occurrence [29].

CONCLUSION

Glypican-3 was expressed in 18% of cases of chronic hepatitis C, 35.1% of cirrhosis and 36.4% of cirrhosis adjacent to carcinoma. Our results suggest that GPC-3 may be considered as an early marker of liver carcinogenesis. 8-OHdG was observed in all cases of chronic hepatitis C, with high labeling index in 82% of cases and low index in 18% of cases. Our study showed high labeling index of 8-OHdG in 52% of cases of HCC. These results indicate that oxidative DNA damage is common in chronic liver disease suggesting a possible link between chronic inflammation and hepatocarcinogenesis. Early detection, continuous monitoring, and subsequent treatment of OS in HCV-related liver disease may prevent further liver damage and slow down or prevent HCC development.

REFERENCES


خفيفة: في مصر، سرطان الخلايا الكبدية هو ثاني أكثر أنواع السرطان شيوعا لدى الرجال وسادس أكثر أنواع السرطان شيوعا عند النساء. مصر لديها أعلى معدلات انتشار التهاب الكبد الوبائي الفيروسي (سي) في العالم وانتشار سرطان الكبد في تزايد في السنوات الأخيرة. الجليبيكاني-3 وهو عضو في البروتينات الريبوبلازمية، يلعب دوراً في نمو الخلايا، والتمايز، والتهرب. 8-هيدروكسيداي اوكسي جوانوسين هو الحمض النووي المؤكد. ويعتبر علامة نقدية في تدفق الحمض النووي من النطف الناجم عن الإجهاد التأكسدي.

الهدف: أجريت هذه الدراسة لتحقيق أظهار الجليبيكاني-3 و 8-هيدروكسي دي أوميغاي جوانوسين في التهاب الكبد الوبائي الفيروسي المزمن (سي) و سرطان خلايا الكبد.

الطريقة والوسائل: تم فحص 50 حالة من حالات التهاب الكبد الوبائي الفيروسي المزمن (سي) و 35 من سرطان خلايا الكبد بواسطة الكيمياء المناعية التسلسلية باستخدام الأجسام المضادة ضد الجليبيكاني-3 و 8-هيدروكسي دي أوميغاي جوانوسين.

النتائج: لوحظ أظهار الجليبيكاني-3 في 18٪ من حالات التهاب الكبد المزمن، وكانت جميع الحالات الإيجابية في الدرجة العالية، بينما أظهار الجليبيكاني-3 في 88٪ من حالات سرطان خلايا الكبد. وقد لوحظ ارتفاع مؤشرل-8-هيدروكسي دي أوميغاي جوانوسين في 79.4٪ من حالات التهاب الكبد المزمن المنخفضه وفي 95.2٪ من حالات التهاب الكبد المزمن المركزة، في حين لوحظ انخفاض مؤشره في 8.2٪ من الدرجة المنخفضة لالتهاب الكبد المزمن وفي 49.8٪ من حالات التهاب الكبد المزمن المرتفع. كان هناك فرق ذات دلالة إحصائية في التعبير عن ال-8-هيدروكسي دي أوميغاي جوانوسين بين الدرجة المنخفضة والعليا لالتهاب الكبد المزمن (قيمة الاحتمال هي 0.10) ولم يكن هناك فرق ذات دلالة إحصائية في أظهار الجليبيكاني-3 و 8-هيدروكسي دي أوميغاي جوانوسين بين الدرجات المختلفة من سرطان خلايا الكبد.

الاستنتاج: أظهر أظهار الجليبيكاني-3 في 18٪ من حالات التهاب الكبد المزمن، وقد لوحظ ارتفاع مؤشرل-8-هيدروكسي دي أوميغاي جوانوسين في جميع حالات التهاب الكبد المزمن، مع مؤشر عال في 82٪ من الحالات ومؤشر منخفض في 18٪ من الحالات. وأظهرت دراسات مؤشر عليا أيضا لل-8-هيدروكسي دي أوميغاي جوانوسين في 51٪ من حالات سرطان خلايا الكبد. هذه النتائج تشير إلى أن الحمض النووي الناتج من التلف التأكسدي هو شائع في الكبد مع الإصابة المزمنة مما يشير إلى احتمال وجود صلة بين الالتهاب الكبد المزمن والإصابة بأورام الكبد السرطانية.