

# **NEUROPROTECTIVE EFFECT OF ADIOL AGAINST LITHIUM PILOCARPINE INDUCED-SEIZURES, BEHAVIRAL CHANGES AND COGNITIVE DEFICITESIN MALE MICE**

#### Heba Ahmed Hassan, Dalia Mohamed Abd El motteleb, Nevertyty Mohamed Mahmoud and Mohamed Mohamed Shehata

Faculty of Medicine, Zagazig University, Department of Clinical Pharmacology, Sharkia, Egypt

#### **Corresponding author:**

Heba Ahmed Hassan Email: drhebahassanpharma58@gmai 1.com

#### ABSTRACT

**Background:** ADIOL (5-androsten- $3\beta$ ,  $17\beta$ -diol) is a metabolite of dehydroepinadro-sterone (DHEA) with antioxidant, anti-inflammatory and antiapoptotic effects. **Objectives**: examine the possible neuroprotective effect of ADIOL against lithium pilocarpine (li-pilo) induced-seizures, behavioral changes and cognitive deficits Methods: 120male mice were divided into six groups, each contain 20 mice which subdivided equally to A for behavioral tests and B for biochemical tests. "Vehicle Group I", "Control Group II "li-pilo", Valproate (VPA) group III (17 mg/kg,i.p). "ADIOL group IV" ADIOL 0.5 mg/kg, "ADIOL group V" ADIOL 1.5 mg/kg "ADIOL group VI" ADIOL 3.5 mg/kg. The animals were observed for the occurrence of spontaneous recurrent seizures (SRS). Open field test, dark light box, acceleration rotarod test and T maze spontaneous alternation were performed. Six mice from each group were euthanized 30 days after pilo injection. Their brains were removed for assessment of oxidant/antioxidant status, anti-inflammatory and antiapoptotic parameters Results: li-pilo produced seizures associated with behavioral changes and cognitive deficits, significant decrease in brain superoxide dismutase (SOD) activity and significant elevation in the brain malondialdehyde (MDA), interleukin1ß (IL-1ß), nuclear factor-kappa B (NF-KB) and caspase-3 (cap-3) in relation to vehicle group. VPA suppressed seizures concomitant with improvement of behavioral changes but worsen short term memory, significant increase in brain SOD activity and significant decrease in MDA, IL-1 $\beta$ , NF-KB and cap-3 levels in relation to li-pilo model. ADIOL (0.5 to 1.5 and 3.5 mg/kg) produced dose dependent decrease in seizures associated with significant improvement of behavioral changes and cognitive deficits, significant increase in brain SOD activity and significant decrease in MDA, IL-1β, NF-KB and cap-3 levels in relation to li-pilo model. **Conclusion:** ADIOL protects against li-pilo induced seizures, behavioral changes and cognitive deficits due to antioxidant, anti-inflammatory, antiapoptotic effects.

Key words: ADIOL, lithium pilocarpine, Seizures, valproate.

#### **INTRODUCTION**

pilepsy is a chronic neurological disorder characterized by repeated spontaneous seizures it is often accompanied by behavioral changes and cognitive deficient<sup>[1]</sup>. The currently used antiepileptic drugs provide asymptomatic anti-seizure effect without effective prophylaxis or cure. Furthermore, a wide range of adverse effects are associated with their long their compliance <sup>[2]</sup>.

Therefore, a need for new antiepileptic drug with better compliance is a promising target.

ADIOL is major metabolite of а [3] (DHEA) dehydroepiandrosterone with reported previously antioxidant. antiinflammatory <sup>[4,5]</sup> and anti-apoptotic activities <sup>[6]</sup>. Several studies have also demonstrated the neuroprotective effects for ADIOL in many neuro-degenerative conditions such as Parkinson's disease and 3-NP-induced neurotoxicity in rat <sup>[6,7]</sup>. ADIOL can mediate

limit

term use and

immunosuppressive effect through ER $\beta$ , in which ADIOL bind to ER $\beta$ -C-terminal binding protein (CtBP) pathway leading to modulate the extent and duration of inflammatory responses [8].

Behavioral changes and cognitive deficient associated with seizures may be caused by underlying structural lesions and effect of seizures and epileptic foci which discharge impulses affect brain function<sup>[9]</sup>.

Increased inflammatory mediators (IL-1β, NF-KB) and cap-3 associated with elevation in oxidative stress are observed following epileptogenic insult and are thought to be implicated in seizure development<sup>[10]</sup>, possibly GABA by modulating and glutamate homeostasis<sup>[11]</sup> and blood brain barrier (BBB) breakdown that leads to leakage of albumin and IgG into the brain<sup>[12]</sup>. Albumin via activation of the transforming growth factor  $\beta$  (TGF)- $\beta$ pathway, leads to increased neural excitability through down-regulation of inward rectifier potassium channel (Kir4.1) and glutamate transporter<sup>[1]</sup>. The current work aimed to study the possible protective effect of ADIOL in versus VPA against li-pilo induced-seizures, behavioral changes and cognitive deficites and to investigate the possible mechanism(s) of action.

# MATERIALS AND METHODS

## Animals

120 adult male Swiss albino mice 8 weeks old weighing 15–35 g were used in the current study. Animals were purchased from the Faculty of Veterinary Medicine, Zagazig University, Egypt. Mice were allowed standard pellet diet and tap water before, during one week of acclimatization period, and through the whole experimental period. They were kept at a constant temperature  $(23 \pm 2^{\circ}C)$ , humidity (60  $\pm 10\%$ ) and a light/ dark (12 h: 12 h) cycle. The animals were randomly assigned to experimental groups. Each mouse was used only once and all tests were performed between 8.00 and 15.00 h. The experiment was performed in the pharmacology department laboratory, faculty of medicine, Zagazig University. Experimental design and animal handling were performed in accordance with protocols approved by the local experimental ethics committee guidelines of the Egyptian Society of Neuroscience, the Ethical Committee

of the Faculty of Medicine, Zagazig University, for Animal Use and the guidelines of the US National Institutes of Health on animal care.

## **Experimental protocol**

Induction of spontaneous recurrent seizures (SRS) by repeated low dose pilocarpine protocol (RLDP)<sup>[13]</sup>, Li.(127 mg/kg,i.p.) injected one day before pilo, Followed by injection of Methylscopolamine bromide (1 mg/kg, i.p.) (30 min) directly before administration of pilo, then the mice received repeated injections of pilo. (10 mg/kg, i.p) every 30 min until they developed convulsive seizures, at stage 4 or 5 according to <sup>[14]</sup>. Stage 4 of seizures was characterized by turn over into side position, and clonic-tonic seizures. In stage 5, rats turned into back position, and showed generalized clonic-tonic seizures. Thus, in both stages, there appeared clonic-tonic convulsions. The animals received up to four injections of pilo.(maximum), or until they developed SE. SE was blocked with administered of xylazine (0.1mg/kg, i.m.)<sup>[15]</sup>. All experimental animals received injections of 5 ml 0.9% NaCl (i.p), directly after SE and twice on the following day after SE to prevent dehydration. The percentage of animals at each seizure stage was calculated in each group <sup>[16]</sup>. Subsequently, the behavior of animals was videotaped and analyzed 24 h a day (7days a week, for 2weeks, beginning 2weeks after pilocarpine injection), for the occurrence of spontaneous recurrent seizures (SRS). After this time, li-pilo model animals were considered epileptic if it presented at least one episode of SRS during the whole observation period <sup>[17]</sup>.

## A-Behavioral tests

4 Weeks after pilo injection we did behavior tests with 1week separate from each test. Open field test for assessment of exploration, dark light box for assessment anxiety, acceleration rotarod test for assessment motor coordination function and T maze spontaneous alternation for assessment working memory.

## 1-Open field test

A rectangular box (30x50 x50 cm), whose floor was divided into 9 small rectangles. Each animal was released in the center of the arena and allowed to explore it for 5 minutes. The number of line crosses and the frequency of rearing are usually used as measures of locomotor activity (number of squares crossed), exploration and anxiety. Grooming behavior (licking of body or paw)<sup>[18]</sup>. Defecation and urination are often used as measures of anxiety <sup>[19]</sup>.The open field was uncontaminated with a 5% water-alcohol solution before behavioral testing to eradicate possible bias due to smells left by previous mice.

## 2-Dark light box test

The test apparatus consists of a box divided into a small (one third) dark chamber and a large (two thirds) brightly illuminated chamber. The box is measured ( $42 \times 21 \times 25 \text{ cm}$ )<sup>[20]</sup>. A restricted opening 3 cm high by 4 cm wide connects the two chambers. A mouse is allowed to move freely between the two chambers for 5 min <sup>[21]</sup>. After each trial, both chambers are cleaned with70% ethanol from defecation and urination of mice. The first latency to enter the dark compartment and the total time spent in lit compartment are indices for anxiety in mice. Transitions are index of exploration.

## **3-Accelerating rotarod test:**

The mice were placed on the already revolving beam facing the opposite orientation so that forward locomotion was necessary in order to avoid a fall. The latencies before falling for four trials per day for 3 days, each trial lasting 1 min, with a 10-min intertrial interval <sup>[22]</sup>. The latency to fall was measured by the Rotarod timer.

## **4-T-maze spontaneous alternation**

Each animal was placed in the starting arm with a sliding door blocking the access to the choice arms. The trials started when the experimenter lifted the door and ended when the animal made a choice and was blocked in one of the arms; entrance was recorded when the animal entered, all four paws, in the selected arm. The first trial was a forced-choice trial where either one of the arms was blocked. The subsequent 14 trials were free-choice trials with both arms open and the mouse was allowed to choose. After a choice, the animal was confined in the selected arm until it made contact with the closed door, which marked the end of the trial. Animals had a maximum time of 2 minutes to perform each trial; after that the animal was gently pushed to the start arm. The T-maze was cleaned with alcohol at 25% between trials to avoid the presence of odour cues. The latency of first entry on the arm and the percentage of alternation were manually recorded <sup>[23]</sup>.

## **B-Biochemical assays**

Six mice from each group were euthanized by decapitation 30 days after pilo injection <sup>[16]</sup>. Their brains were quickly removed in liquid nitrogen. The whole brain was homogenized in ice-cold saline for estimation of SOD, MDA, IL- $1\beta$ , NF- $\kappa$ B, cap-3 were estimated utilizing enzyme linked immunosorbant assay (ELISA) kits purchased from (mybiosource, USA), (life spane bioscience, USA),(CUSABIO Bender Med Systems, USA), (EIAab, USA) and (CUSABIO, USA), respectively and results were expressed as Microgram/Gram(u/g) for SOD, picograms per gram tissue (pg/g tissue) for IL- $1\beta$ , NF- $\kappa$ B and cap-3 and as nanograms per gram tissue (ng/g tissue), for MDA level.

## **Statistical Analysis**

The obtained Continuous variables were tabulated as means ± SEM. Comparison between different groups were made using one way analysis of variances (one-way ANOVA) followed by Post-Hoc (least significant difference "LSD") tests as described by Armitage and Berry <sup>[24]</sup>. The categorical expressed as variables were number а percentage. Percent of categorical (ordinal variables were compared using Chi-square test for trend). The differences were considered to be significant when p < 0.05. All other results are carried by Graph Pad prism software version 5.

## RESULTS

## Pharmacological results

In li-pilo model, administration of VPA and ADIOL (0.5, 1.5 and 3.5 mg/kg) protected 95%, 80%, 95% and 100% of mice respectively from reaching stage 4 seizures (p < 0.05) (Table1).

Tabel (1): Percentage of reduction of stage 4 seizures in	nduced by valproate (17mg/kg, i.p.), ADIOL
(0.5, 1.5 and 3.5g/kg, s.c.) in li-pilo model in mice	

	Control	Valproate	ADIOL 0.5	ADIOL 1.5	ADIOL 3.5
Percentage	15%	95% <sup>a</sup>	80% <sup>a</sup>	95% <sup>a</sup>	100% <sup>ab</sup>

Values are means of 6 mice ±SEM. <sup>a</sup> P <0.05 compared to control group, <sup>b</sup> P <0.05 compared to VPA group

#### **A-Behavioral tests results**

1- Open field(table 2)

Regarding ambulation frequency, Control group produced significant increase (p<0.05) in ambulation frequency in comparison to vehicle. While VPA (17mg/kg) and ADIOL (1.5 and 3.5mg/kg) produced significant increase (p<0.05) in ambulation frequency in comparison to control group. While, ADIOL 0.5 mg/kg was produced insignificant change in ambulation frequency in comparison with control group.

Regarding no of rearing, Control group produced significant decrease (p<0.05) in mean number of rearing when compared to vehicle group. Even so with valproate (17mg/kg) was produced significant increase (p<0.05) in mean number of rearing when compared to control group. While ADIOL (0.5, 1.5 and 3.5mg/kg) were produced insignificantly change (p>0.05) mean number of rearing in the open field test when compared to control group. Regarding number of grooming, control group produced insignificant change (p>0.05) in mean number of when compared to vehicle, But valproate (17mg/kg) and ADIOL (0.5 and 1.5 mg/kg) produced insignificant change (p>0.05) in mean number of grooming when compared to control group and vehicle group. While ADIOL 3.5 mg/kg produced significantly increase (p<0.05) in mean no of grooming as compared to control group but, insignificant change (p>0.05) in mean number of grooming when compared to control group but, insignificant change (p>0.05) in mean number of grooming when compared to control group but, insignificant change (p>0.05) in mean number of grooming when compared to vehicle group.

Regarding number of fecal boli, control group produced significant decrease (p<0.05) in mean number of fecal boli when compared to vehicle. While, valproate (17mg/kg) and ADIOL (0.5, 1.5 and 3.5 mg/kg) produced significant decrease (p>0.05) in mean number of fecal boli when compared to control with no significant change in compared with vehicle group.

Table (2): Effects of VPA (17mg/kg), ADIOL (0.5, 1.5 and 3.5mg/kg) on the mean values ± SEM on exploratory behaviors in open field test (ambulation frequency, no of rearing, no of grooming and no of fecal boli) in the mice li-pilo model.

Group N=6	Vehicle	control	Valproat	ADIOL 0.5	ADIOL 1.5	ADIOL 3.5
			e			
Ambulation	2.8 ±77.3	$25.8^{a} \pm 2.13$	76.5±3	66.6 <sup>a</sup> ±7.87	<b>98.6</b> <sup>a</sup> ±17.8	107.7±14.2
frequency						
No Rearing	38.6±5.8	$7^{a} \pm 2.3$	<b>34.6</b> <sup>b</sup>	17±7.35	17.6±4.85	26.3±7
C			±7.8			
No Grooming	12.6±2.04	7±1.9	8.6±0.98	4.33±0.95	5.6±1.08	26.3 <sup>b</sup> ±7.08
No Fecal boli	2.5±0.34	7 <sup>a</sup> ±1.4 B	2.33 <sup>b</sup>	1.5±0.34	$0.83^{b} \pm 0.4$	0.83 <sup>b</sup> ±0.4
			±1.3			

Values are means of 6 mice ±SEM. <sup>a</sup> P <0.05 compared to vehicle group, <sup>b</sup> P <0.05 compared to li-pilo

# control group

## 2-Dark light box test (table 3)

Regarding number of transition, control group produced significant decrease (p<0.05) in mean number of transition when compared to vehicle group. Also valproate (17mg/kg) and ADIOL (0.5, 1.5 and 3.5mg/kg) could not produce any significant change (p>0.05) in mean number of transition in relation to control group. Also valproate (17mg/kg) and ADIOL (3.5mg/kg) insignificantly (p > 0.05) decrease mean no of transition as compared to vehicle group. While, ADIOL 0.5 and 1.5 mg/kg significantly (p<0.05), decreased mean no of transition in comparison with vehicle group.

Regarding number of time that mice spent in dark compartment in sec, control group produced significant change (p<0.05) in mean dark time

when compared to vehicle group. But valproate (17mg/kg) and ADIOL (0.5, 1.5 and 3.5mg/kg) could not produce any significant change (p>0.05) in mean dark time when compared to control group or vehicle.

Regarding number of time that mice spent in light compartment in sec, control group produced significant decrease (p<0.05) in mean light time when compared to vehicle group. But valproate (17mg/kg) and ADIOL (0.5 and 1.5mg/kg) could not produce any significant change (p>0.05) in mean light time in when compared to control group or vehicle group. Furthermore, ADIOL (3.5mg/kg) had produced significant increase (p<0.05) in mean light time when compared to control group by with no significant change (p>0.05) with vehicle group.

Table (3): Effects of VPA (17mg/kg), ADIOL (0.5, 1.5 and 3.5mg/kg) on the mean values  $\pm$  SE on exploratory behaviors in dark light box test (no of transition, mean dark time and mean light time) in mice lithium pilocarpine model.

Group	vehicle	control	Valproa	ADIOL 0.5	ADIOL 1.5	ADIOL 3.5
			te			
Mean No of	8.6±1.2	$2^{a} \pm 0.5$	$4.3 \pm 0.5$	$2.3^{a} \pm 0.4$	<b>3.6</b> <sup>a</sup> ± <b>0.8</b>	6.3±2
Transition						
Mean Dark Time	171.7±1	272.7 <sup>a</sup> ±5.6	233.3±1	201.7±21.9	218.7±29.3	200.5±21.7
(sec)	2.7		4.4			
Mean Lit Time	30.5±5	4.5±1.8	43.6±9.	38.3±14.8	65.8±27	$61.8^{b} \pm 21.7$
(sec)			3			

Values are means of 6 mice  $\pm$ SEM. <sup>a</sup> P <0.05 compared to vehicle group, <sup>b</sup> P <0.05 compared to li-pilo control group

## **3-Accelerating rotarod test (table 4)**

Regarding the latency to fall from the rotarod bar, control group produce no significant change (p>0.05) in mean mean latency period when compared to vehicle group. While valproate (17mg/kg) significantly (p<0.05) increased mean latency period as compared to control group, with no significant difference (p>0.05) from vehicle group. Furthermore, ADIOL (0.5 and 1.5 mg/kg) was insignificantly (p>0.05) decreased mean latency to fall from as compared to control group, with no significant difference (p>0.05) from vehicle group. Also ADIOL (3.5 mg/kg) significantly (p<0.05) increased mean latency period in comparison to control group.

Table (4): Effects of VAP (17mg/kg) and ADIOL (0.5, 1.5 and 3.5mg/kg) on the mean values  $\pm$  SE on mean latency to fall in sec in rotarod test (3 days) in mice lithium pilocarpine model.

Group	vehicle	control	valproate	ADIOL 0.5	ADIOL 1.5	ADIOL 3.5
Mean latency To fall in sec	39.6±3.6	25.2±3.5	$55.3^{b} \pm 4.1$	33.5±2.54	38.9±2.7	52.9 <sup>b</sup> ±5.9

Values are means of 6 mice ±SEM. <sup>a</sup> P <0.05 compared to vehicle group, <sup>b</sup> P <0.05 compared to li-pilo control group

# **4-T-maze** spontaneous alternation (table 5)

Regarding spontaneous alternation, control group produced significant decrease (p<0.05) in % of spontaneous alternation when compared to vehicle group. While, valproate (17mg/kg) were insignificantly (p>0.05) increased the mean % of spontaneous alternation as compared to control group, it is still insignificantly (p>0.05) lower than vehicle group. Also ADIOL (0.5and 1.5 mg/kg) were insignificantly (p>0.05) increased mean % of spontaneous in relation to control group. These were insignificantly (p>0.05) lower than vehicle group. While, ADIOL (3.5 mg/kg) was significantly (p<0.05) was mean % of

spontaneous with control group and It was insignificantly (p>0.05) lower than vehicle group.

Regarding the Latency in the starting arm, control group produced significant decrease (p<0.05) in mean Latency in the starting arm when compared to vehicle. Valproate (17mg/kg) produced significant (p<0.05 decrease in mean Latency in the starting as compared control group, it is still insignificantly (p>0.05) lower than vehicle group. Also ADIOL (0.5, 1.5 and 3.5 mg/kg) were significantly (p<0.05) decreased mean Latency in the starting arm in comparison with control group. These were significantly (p<0.05) higher than vehicle group.

Table (5): Effects of VAP (17mg/kg) and ADIOL (0.5, 1.5 and 3.5mg/kg) on the mean values ± SE on % of spontaneous alternation and latency to first entry in start arm in spontaneous alternation T-MAZE test in mice lithium pilocarpine model.

Group	vehicle	control	valproate	ADIOL 0.5	ADIOL 1.5	ADIOL 3.5
% Spontaneous alternation	90.9±2. 3	61.8 <sup>a</sup> ± 13.1	77± 5.4	88.9±5	86.4±1.9	90.2 <sup>b</sup> ±3.8
Latency to first Entry in start Arm	10.8±1. 2	23.83 <sup>a</sup> ±4.9	12.5 <sup>b</sup> ±0.7	12.1 <sup>b</sup> ±0.8	17.6 <sup>b</sup> ±4.85	10.8 <sup>b</sup> ±0.6

Values are means of 6 mice ±SEM. <sup>a</sup> P <0.05 compared to vehicle group, <sup>b</sup> P <0.05 compared to li-pilo control group

## **Biochemical results**

Compared to vehicle animals, li-pilo mice produced significant reduction (p<0.05) in brain SOD activity by 84.2%. Moreover, epileptic mice showed significant (p<0.05) increase in brain level of MDA, IL-1 $\beta$ , NF- $\kappa$ B and cap-3 by 547.3%, 459.2%, 860% and 522.2%, respectively (figure1: A,B,C,D,E). Mice which pretreatment with VPA for 2 days before pilo produced significant (p <0.05) increase in brain SOD activity by 373.3%, Furthermore, VPA significantly (p<0.05) decreased brain level of MDA, IL-1β, NF-KB and cap-3 by 78%, 76.1%, 79.3% and 72.9%, respectively as compared to li-pilo control group(figure1A,B,C,D,E). ADIOL(0.5mg/kg) for 2 days before pilo produced significant (p < 0.05) increase in brain SOD activity by 66.6%, also produce significant (p<0.05) decrease in brain level of MDA, IL-1 $\beta$ ,

NF-KB and cap-3 by 36.5%, 41.7%, 37.5% and 38.7%, respectively as compared to li-pilo control group (figure1A,B,C,D,E). While with pretreatment epileptic of mice with ADIOL(1.5mg/kg) for 2 days before pilo produced significant (p <0.05) increase in brain SOD activity by 166.6% associated with significant (p<0.05) decrease in brain MDA, IL1β, NF-κB and cap-3 by 51.4%, 55.6%, 56.3% and 54.2%, respectively as compared to li-pilo (figure1A,B,C,D,E). control group with increasing ADIOL dose of to (3.5)mg/kg), animals produced significant (p < 0.05) increase in brain SOD activity by 300% concomitant with significant (p<0.05) decrease in brain MDA, IL-1 $\beta$ , NF-KB and cap-3 by 67.6%, 65.5%, 73.3% and 69.8%, respectively as compared li-pilo control to group(figure1A,B,C,D,E)



**Fig. 1:** showing comparison between different groups regarding brain SOD, MDA, IL-1 $\beta$ , NF-KB and cap-3.Values are means of 6 mice ±SEM. @ P <0.05 compared to vehicle group, \$P <0.05 compared to li-pilo control group, \*P <0.05 compared to valproate group, =P <0.05 compared to ADIOL 0.5 group, #P <0.05 compared to ADIOL 1.5 group.

#### DISCUSSION

The results of the current study revealed that 85% of mice subjected to RLDP reached stage 4/5 clonic convulsions and were associated with significant tissue reduction in SOD activity with significant increase in MDA level in brain mice. These results are in agreement with those of Ahmad et al<sup>[25]</sup>; Armenta et al. <sup>[26]</sup>, who reported high levels of MDA and low SOD activity in brain of epileptic rate which induced by li-pilo.

It is postulated that oxidative stress resulting from excessive free-radical release is likely implicated in the initiation and progression of epilepsy <sup>[27,28]</sup> Oxidative stress occurs during epileptogenesis due to seizures-associated glutamate excitotoxicity and NMDA receptor <sup>[29]</sup> leading to increase in overactivation oxidation of macromolecules of the neurons just before the neuronal loss <sup>[30]</sup>. In its turn, free radicals inhibit glutamine synthase (which transforms glutamate into glutamine inside astrocytes), inhibit glutamate decarboxylase (which converts glutamate into GABA) and inactivate glutamate transporter causing increase glutamate concentration and diminish GABA level <sup>[31]</sup>. Also, ROS may activate NFkB, leading to the production of pro-inflammatory cytokines such as IL-1, IL-6, and TNF- $\alpha$ , which in turn enhance inflammation and, therefore, the generation of more reactive species <sup>[32.33]</sup>.

In the current study, i.p injection of li-pilo significantly elevated brain level of proinflammatory cytokines; IL1 $\beta$  and NF- $\kappa$ B levels. These findings are in agreement with Yang et al. <sup>[34]</sup> who found that serum levels of IL-1 $\beta$ , NF- $\kappa\beta$  and TNF- $\alpha$  in children with MTLE were significantly higher than the vechile as well as in animal models of epilepsy induced by chemoconvulsants or by electrical stimulation <sup>[35,36,37]</sup>.

The possible explanation could be seizure activation caused neuronal cell loss and the destruction of the BBB [38], associated with increasing inflammatory responses and neural excitability in the brain <sup>[39]</sup>. IL-1β and NF-KB produced by astrocytes leads to the activation of the PI3K/Akt/mTOR signaling pathway after combining with the interleukin-1 receptor (IL-1RI) on the neurons, and that the activation of this signaling pathway plays an important part in pathogenesis of MTLE<sup>[40]</sup>. cytokines the contribute to epileptognesis in multiple ways, by increasing neuronal hyperexcitability via astrocyte-mediated blocking reuptake of glutamate from the synaptic space <sup>[35]</sup>, also enhance NMDA receptor function<sup>[41]</sup> with alternation of GABAergic neurotransmission<sup>[42]</sup>. Also modulates voltage-gated ion channels and

[43] to channelopathies NF-κB contributes leads to activation activation of nonpathway [44] which transcriptional cause phosphorylation of voltage-dependent and receptor-coupled ion channels, thereby directly affecting neuronal excitability and seizure threshold [36].

present study showed significant The elevation of caspase 3 level in brain mice which injected with li-pilo. This work coincided with Khadrawy et al. <sup>[45]</sup> who reported elevation of cap3 level in rat brain after induction of SE by lipilo. Fujikawa <sup>[46]</sup> suggested that SE leads to excessive activation of NMDA receptors by glutamate, allowing calcium influx through their cation receptor-operated channels causing excitotoxicity <sup>[47]</sup> resulting in downstream swelling and rupture of intracellular organelles and activated proteolytic enzymes leading to cell death  $^{[48]}$ . NF- $\kappa$ B activation with epilepsy can also induce the generation of neurotoxic free radicals; resulting in neuronal apoptosis <sup>[49]</sup>. The active caspase 3 is responsible for the cleavage of key intracellular structural and survival proteins and activates the enzyme responsible for the DNA fragmentation<sup>[50]</sup>.

The Present study shows, VPA administration protects against li-pilo induced SE with reducing brain oxidative stress, hampered brain level of inflammatory mediators and protect brain cell from apoptosis. According to Rajeshwari et al. <sup>[51]</sup>, the antioxidant effect of VPA is attributed to its free radical scavenging ability, enhancement of enzymatic antioxidant network (SOD, catalase and glutathione peroxidase). In addition, VPA was reported to elevate the level of nonenzymatic antioxidants namely, vitamin C, vitamin E, and reduced glutathione, which scavenge the residual free radicals escaping from decomposition by the antioxidant enzymes <sup>[52]</sup>. VPA block NMDA receptor and Na<sup>+</sup> channelmediated glutamate release and excitotoxicity in cerebral nerve endings <sup>[53]</sup> which may be an additional contributing factor in ROS production inhibition. As a HDACs inhibitor, VPA has the ability to activate the expression of genes targeting antioxidant enzymes (e.g. SOD), and induce several neuroprotective proteins such as angiogenin, brain-derived neurotrophic factor (BDNF), endothelial cell growth factor (ECGF1), and ganglia cell-derived neurotrophic

<sup>[54,55]</sup>The anti-inflammatory factor (GDNF) effect of VPA is due to modulation of ion channels (Na<sup>+</sup>, K<sup>+</sup>, Ca<sup>2+</sup> and Cl<sup>-</sup>) in immune cells leading to suppression of cytokines production <sup>[56,33]</sup>. **Himmerich et al.** <sup>[33]</sup>, reported that the cytokine production is due to elevation in ROS generation. Thus, the anti-inflammatory effect of valproate may be due to antioxidant effect. VPA exhibits dose-dependent biphasic effect on apoptosis of activated lymphocytes probably through differential modulation of several apoptosis-related signaling pathways as VPA in low dose had anti-apoptotic effect through activation of extracellular signal-regulated kinases (ERK) pathway and phosphorylation of anti-apoptotic molecules such as Bcl-2<sup>[57]</sup> associated with significant elevation of phosphorylated STAT3 lead to the attenuation of activation-induced apoptosis, but at high doses it promoted apoptotic cell death by induction of p38 mitogen-activated protein kinase (MAPK) and mitochondrial apoptosis pathway with increase dephosphorization of STAT3 resulting in enhanced activation-induced apoptotic cell death [58,59]

This work demonstrated that, pretreatment of epileptic mice with ADIOL produced significant dose dependent antiepileptic effect associated also with dose dependent antioxidant, antiinflammatory and antiapoptotic effects.

These results are matched with Reddy and Jian <sup>[60]</sup>; Carver and Reddy <sup>[61]</sup> they stated that ADIOL caused dose-dependent suppression of seizures in mouse hippocampus kindling epilepsy. This could be explained by Clossen and Reddy <sup>[62]</sup>, who stated that, ADIOL (a neurosteroid) induced changes in neuronal excitability, through modulation of GABAARs (<sup>[63]</sup>which permit influx of chloride ions into the cell causing hyperpolarizing of the membrane and reduced excitability at cellular and systems level <sup>[64]</sup>. As ADIOL at low concentrations, act as positive allosteric modulator, potentiating Cl<sup>-</sup> influx by increasing the duration and frequency [60,61] of channel opening but at higher concentrations, can directly activate the GABAARs<sup>[65,61]</sup>

In agreement with our study, **Hanna et al**. <sup>[6]</sup> reported that ADIOL (25 mg/kg, s.c.) for two days before 3- 3-nitropropionic acid displayed antioxidant, anti-inflammatory and antiapoptotic

activities as evidenced by the elevation of cortical and striatal reduced glutathione levels, reductions of cortical and striatal MDA, striatal TNF- $\alpha$  and IL-6 levels. Only a small number of iNOS and caspase-3 positive cells were detected in sections from rats pretreated with ADIOL. Also **Salama et al**. <sup>[7]</sup> stated that ADIOL in doses 0.35, 3.5 and 35 mg/kg decreased striatal and nigral NF- $\kappa$ B level in rotenone induced-parkinson's disease rat model with reduced cap3, Bax, Bax/Bcl level. Also ADIOL inhibited the expression of IL-1 $\beta$  upon LPS stimulation of human microglia and the expression of iNOS mRNA in mouse astrocytes when cells were stimulated by IL-1 $\beta$ <sup>[67]</sup>.

**Saijio et al.** [67] explained that ADIOL functions as a selective modulator of  $(ER\beta)$  to suppress inflammatory responses of microglia and astrocyte by mediate recruitment of ERB-Cterminal binding protein (CtBP) corepressor complexes to Activator protein-1which is transcriptional factor responsible for gene expression for cytokines and growth factors<sup>[68]</sup>., leading to suppressing genes that amplify inflammatory responses and activate Th17 T cells. Reduction of endogenous ADIOL or ERB expression results in exaggerated inflammatory responses to TLR4 agonists. ADIOL promote experimental CtBP recruitment prevents autoimmune encephalomyelitis in an ERβdependent manner. These findings provide evidence for an ADIOL/ER<sub>β</sub>/ CtBPtransrepression pathway that regulates inflammatory responses in microglia and can be targeted by selective ER $\beta$  modulators <sup>[67]</sup>.

In both O.F and LDT, control group exhibited decrease in exploratory behavior with increase behavior in relation to anxiety vechile group. These results are consistent with [69,70] who reported that, reduced locomotor and exploratory behavior associated with increased in anxiety behavioral in li-pilo model, was observed in the Irwin screen, open field, hole-board, light-dark box. As SE induced by li-pilo causes extensive neuronal loss in the hippocampus, cortex, amygdala and thalamus <sup>[71]</sup>. These structures are known to be functionally important in anxiety and exploration <sup>[72,73]</sup>, also damage to them produce behavioral impairment.

But pretreatment of mice with VPA, animals increase in exploratory behavior and decrease in

anxiety behavior but in LDT no difference was detect in the behavior of mice in relation to control group. In agreement with current study **Zádori et al.** <sup>[74]</sup> stated that VPA significantly increased the ambulation distance (no of crossed square) and increase in spontaneous locomotor activity in open field test in in a transgenic mouse model of Huntington's disease. Due to VPA increases GABA levels in the substantia nigra pars reticulate which is an important part of both direct and indirect pathways of the basal ganglia circuitry leading to decrease inhibition of the thalamus, resulting in increased locomotion

With pretreatment of mice with ADIOL 0.5 mg/kg exhibited no effect on exploratory behavior but significant decrease in anxiety behavior while with increase dose to 1.5 and 3.5mg/kg animal exhibit significant increase in exploratory behavior and significant decrease in anxiety behavior. But in LDT no difference was detect in the behavior of mice in relation to control group except for ADIOL 3.5 mg/kg decrease anxiety behavior. In line with our [7] Salama et al. results. showed that pretreatment of rotenone-induced Parkinson disease rat model rate by ADIOL 3.5mg/kg increase in no of crossed squares also ADIOL 3.5 and mg/kg showed significant 35 improvement in the exploratory and increase in rearing frequency in rat due to increased DA level with significant decline in DA turnover. Also, **Tsutsui and Haraguchi** <sup>[75]</sup> reported the modulatory effects of neurosteroids on DA in pineal body which stimulates locomotor behavior. Also ADIOL is a selective modulator of ERβ<sup>[67]</sup>. **Borrow and Handa**<sup>[76]</sup>reported that ERB had anxiolytic effect which observed in pharmacological studies utilizing selective ERB agonists as diarylpropionitrile caused decreased anxiety behavior in rats and mice in a number of tests, including the open field test, elevated zero maze test elevated plus maze test, and light-dark box test <sup>[77,78]</sup>. As ER increased the expression of tryptophan hydroxylase (TPH) (the rate-limiting enzyme in serotonin synthesis), this accompanied by increases in the levels of TPH enzymatic activity and total serotonin which act on 5-HT1A receptor leading to anti-anxiety effect<sup>[79]</sup>.

In this study our observations regarding the rotarod test performance show that there is no

significant difference in latency to fall, between li-pilo group and vechile group. This result coincides with wu et al. <sup>[80]</sup> who reported that lipilo model had no effect on motor coordination rotarode test).This (no effect on work VPA produce significant demonstrates that improvement in motor coordination in comparison with li-pilo group. In agreement with our result Weihl et al.<sup>[81]</sup> who suggested that VPA improve muscle strength and function in patients with type III/IV spinal muscle atrophy by increased transcription of spinal muscular atrophy genes and protein in vitro leading to increase in quantitative muscle strength and subjective function of muscle. Also, **Yu et al.**<sup>[82]</sup> demonstrated the ability of VPA to improve motor coordination in mice with traumatic brain injury by inhibiting glycogen synthetase kinase 3 (GSK-3) and HDAC effect of VPA leading to reduction of lesion volume and attenuation of blood-brain barrier disruption. Furthermore, Esteves et al <sup>[83]</sup> reported that chronic VPA (200mg/kg) treatment of Mouse Model of Machado-Joseph Disease lead to improvement in the motor performance, given by the beam balance, motor swimming, rotarod due to increase in histone acetylation by its HDACi effect which mediate gene transcription leading to improve ataxic symptoms in a transgenic mouse<sup>[84]</sup>. ADIOL 0.5 and 1.5 mg/kg produce significant improvement in no motor coordination in comparison with control group but with increasing the dose to 3.5 mg/kg showed significant improvement in motor in comparison to li-pilo group.As ADIOL is a selective modulator of  $(ER\beta)$  that had significant effect on motor coordination due to its physiological role in cerebellum of adults <sup>[85]</sup> by preventing neuronal deaths, inducing formation and increasing information of synapses transmission through nerve impulses <sup>[86]</sup>, maintaining the structure <sup>[87]</sup> and function of cerebellum<sup>[86]</sup>.

With i.p injection of li-pilo to mice, animal showed impairment in memory. These results are agreed with **Hoppe et al.**<sup>[88]</sup> who stated that patients with MTLE often associated with memory impairment. Also impaired learning and memory is observed in the pilo model of MTLE in rats and mice <sup>[89]</sup>. Cognitive and behavioral impairment that associated with TLE are resulting from hippocampal sclerosis, Loss of neurons in the hippocampus and associated gliosis<sup>[90]</sup>

While pretreatment of epileptic mice with VPA, animals show no improvement in memory impairment. Some studies coincide with currant work, as VPA and other anticonvulsant mood stabilizers have generally been found to have some adverse effects on cognition in patients with epilepsy <sup>[91]</sup>. **Sgobio et al.** <sup>[92]</sup>, reported that VPA induced morphologic alterations and impairment in specific hippocampal-dependent memory task as VPA inhibits acetylcholine esterase (AchE) causing increase acetyl choline in barin leading to cognitive deficits <sup>[93]</sup>

In present model, pretreatment with ADIOL 0.5 and 1.5 mg/kg show no improvement in memory impairment but with increasing dose of ADIOL to 3.5 mg/kg produce significant improvement in memory impairment that caused by li-pilo injection. This could be explained by **Bodo and Rissman** <sup>[94]</sup> who reported the role of ER  $\beta$  in memory ability that vary along the menstrual cycle in women <sup>[95,96]</sup> and decline in this ability by post-menopausal women which counteracted by estrogen replacement therapy<sup>[97]</sup>.

# CONCLUSION

ADIOL protects against li-pilo induced seizures, behavioral changes and cognitive deficits due to its antioxidant, anti-inflammatory, antiapoptotic effect.

## ABBREVIATIONS

ADIOL: 5-androsten- $3\beta$ ,  $17\beta$ -diol; AEDs: antiepileptic drugs; AKT: Protein kinase B (PKB);ANG: Angiogenin; BBB: blood brain barrier; BDNF: Brain-derived neurotrophic factor; cap 3: caspase3; CtBP: ERβ-C-terminal binding protein; DHEA: dehydroepinadrosterone;ECGF1: Endothelial cell growth factor; ELISA: enzyme linked immunosorbant assay; ER stress: endoplasmic reticulim stress ;ERK: Extracellular receptor kinase; ER $\beta$ : estrogen receptor beta; GABA: gamma amino butyric acid; GABAAR: gamma amino butyric acid receptor; GDNF: Glial cellderived neurotrophic factor; GSH: Reduced glutathione; HDAC: histone deacetylase activity; i.m.: intramuscular; i.p: intraperitoneal; I/R: Ischemia reperfusion IgG: immunoglobulin g; IL-1:interleukine 1;IL-1R: IL-1 receptor;IL1 $\hat{\beta}$ : inteleukin1<sub>β</sub>;IL-6: interleukine 6;Kir4.1: inward rectifier potassium channel; li-pilo: lithium pilocarpine LPS:lipopolysaccharides ;MAPK: mitogen-activated protein kinase :MDA: malondialdehyde; MES: maximal electroshock seizures; MTLE: mesial temporal lobe epilepsy; MTOR; mammalian target of rapamycin; n: number of animals; NF-KB: nuclear factor kappa b; NMDA: N-methyl-D-aspartic acid; ROS: Reactive species;PI-3-K: oxygen Phosphoinositide-3-kinase; PTZ: Pentanyltetrazol ; RLDP: repeated low dose pilocarpine protocol; s.c.: subcutaneous; SOD: superoxide dismutase; SRS: spontaneous recurrent seizures; TGF-<sub>β</sub>: transforming growth factor  $\beta$ ;TLR : Toll like receptor: TNF- $\alpha$ : Tumor Necrosis Factor-α;VPA: valproate.

## REFERENCES

- 1. Shimada T., Takemia T., Sugiura H. and Yamagata K. role of inflammatory mediators in pathogenesis of epilepsy. Mediators of inflammation. 2014; V 2014.
- 2. Cevik B, Solmaz V, Aksoy D, Erbas O. Montelukast inhibits pentylenetetrazolinduced seizures in rats. Med Sci Monit. 2015;21:869-874
- 3. Simard J., Ricketts M.L., Gingras S., Soucy P., Feltus F.A. and Melner M.H. Molecular biology of the 3beta-hydroxysteroid dehydrogenase/delta5-delta4 isomerase gene family. Endocr Rev. 2005; 26; 525–582.
- Auci, D., Nicoletti, F., Mangano, K., Pieters, R., Nierkens, S., Morgan, L., Offner,H., Frincke, J. and Reading, C. Anti-inflammatory and immune regulatory properties of 5androsten-3beta, 17beta-diol (HE2100), and synthetic analogue HE3204: implications for treatment of autoimmune diseases. Ann. N. Y. Acad. Sci.2005; 105; 730-742.
- 5. Nicoletti F., Auci D. L., Mangano K., Flores-Riveros J., Villegas S., Frincke J. M. et al. 5-Androstenediol Ameliorates Pleurisy, Septic Shock, and Experimental Autoimmune Encephalomyelitis in Mice. Autoimmune Diseases.2010; volume (2010).757432.
- 6. Hanna, D. M., Tadros M.G. and Khalifa A.E. ADIOL protects against 3-NP-induced neurotoxicity in rats: Possible impact of its antioxidant, anti-inflammatory and anti-apoptotic actions. Prog Neuropsychopharmacol Biol Psychiatry. **2015**; 60; 36-51.
- 7. Salama, R. M., Tadros M.G., Schaalan M.F., Bahaa N., Abdel-tawab A.W. and Khalifa A.E. Potential neuroprotective effect of

androst-5-ene- $3\beta$ ,  $17\beta$ -diol (ADIOL) on the striatum, and substantia nigra in Parkinson's disease rat model. Journal of Cellular Physiology.**2018**; 233(8); 5981-6000.

- 8. Gosselin D. and Rivest S. : Estrogen receptor transrepresses brain inflammation Cell. 2011; 145(4); 495-497
- 9. **Berg T.A.** Epilepsy, cognition, and behavior: The clinical picture. Epilepsia (**2011**); 52; 7–12.
- 10. Mattson M. P. and Meffert M. K. Roles for NF-kappaB in nerve cell survival, plasticity, and disease. Cell Death Differ.2006; 13(5); 852-860.
- Postnikova T. Y., Zubareva O. E., Kovalenko A. A., Kim K. K., Magazanik L. G. and Zaitsev A. V. Status epilepticus impairs synaptic plasticity in rat hippocampus and is followed by changes in expression of NMDA receptors. Biochemistry (Mosc). 2017; 82; 282– 290.
- 12. Van Vliet E. A., Aronica E. and Gorter J. A. Role of blood-brain barrier in temporal lobe epilepsy and pharmacoresistance. Neuroscience.2014; 277; 455-473.
- SzyndlerJ., Wierzba-Bobrowicz T., Skórzewska A., Maciejak P., Walkowiak J., Lechowicz W., Turzyńska D., Bidziński A. and Płaźnik A. Behavioral, biochemical and histological studies in a model of pilocarpineinduced spontaneous recurrent seizures. Pharmacology Biochemistry and Behavior. 2005; 81(1): 15-23.
- Becker A., Grecksch G., Thiemann W. and Hoellt V. Pentylenetetrazol-kindling modulate stimulated dopamine release in the nucleus accumbens. Pharmacol Biochem Behav .2000; 66; 425–8.
- 15. Sinclair M. D. A review of the physiological effects of  $\alpha$ 2-agonists related to the clinical use of medetomidine in small animal practice. The Canadian Veterinary Journal.2003; 44(11); 885–897.
- 16. **Abdallah D.M.** Anticonvulsant potential of the peroxisome proliferator-activated receptor  $\gamma$  agonist pioglitazone in pentylenetetrazole-induced acute seizures and kindling in mice. Brain Res. **2010**; 1351:246-253.
- 17. Glien M., Brandt C, Potschka H., Voigt H., Ebert U. and Loscher W. Repeated low-dose treatment of rats with pilocarpine: low mortality but high proportion of rats developing epilepsy. Epilepsy Res.2001; 46(2); 111-119.
- 18. **Espejo E.F.** Effects of weekly or daily exposure to the elevated plus-maze in male mice. Behavioural Brain Research. **1997**; 87; 233-238.
- 19. **Hall C.S.** Emotional behavior in the rats Defecation and urination as measures of

individual differences in emotionality. Journal of Comparative Psychology.**1934**; 18(3); 385-403.

- Crawley J. and Goodwin F. K. Preliminary report of a simple animal behavior model for the anxiolytic effects of benzodiazepines. Pharmacol Biochem Behav.1980; 13(2); 167-170.
- Costall B., Jones B. J., Kelly M. E., Naylor R. J. and Tomkins D. M. Exploration of mice in a black and white test box: Validation as a model of anxiety. Pharmacol Biochem Behav.1989; 32(3); 777-785.
- 22. Lalonde R., Hayzoun K., Selimi F., Mariani J. and Strazielle C. Motor coordination in mice with hotfoot, Lurcher, and double mutations of the Grid2 gene encoding the delta-2 excitatory amino acid receptor. Physiol Behav. 2003; 80(3); 333-9.
- 23. **Deacon R.M. and Rawlins J.N.** T-maze alternation in the rodent. Nat Protoc. **2006**; 1(1); 7-12
- 24. **Armitage P, Berry G.** Statistical methods in medical research, 3<sup>rd</sup> edition; pp 620. Blackwell Scientific Publications, Oxford. London, **1994**.
- 25. Ahmad M., Abu-Taweel G.M., Aboshaiqah A.E. and Ajarem J.S.The Effects of Quinacrine, Proglumide, and Pentoxifylline on Seizure Activity, Cognitive Deficit, and Oxidative Stress in Rat Lithium-Pilocarpine Model of Status Epilepticus, Oxidative Medicine and Cellular Longevity,2014; vol. 2014, Article ID 630509, 11 pages
- 26. Armenta M. M., Ruíz C. N., Rebollar D. J., Martínez E. R. and Gómez P.Y. Oxidative Stress Associated with Neuronal Apoptosis in Experimental Models of Epilepsy. Oxidative Medicine and Cellular Longevity, **2014**; vol. 2014, Article ID 293689, 12 pages.
- 27. Diniz T. C., Silva J. C., de Lima-Saraiva S. R. G., Ribeiro F. P. R. de A., Pacheco A. G. M., de Freitas R. M. et al. The Role of Flavonoids on Oxidative Stress in Epilepsy. Oxidative Medicine and Cellular Longevity.2015; volume 2015; article ID171756.
- 28. Geronzi U., Lotti F. and Grosso S. Oxidative stress in epilepsy. Expert Review of Neurotherapeutics.2018; 18(5); 427-434.
- 29. Shi L.L., Dong J., Ni H., Geng J. and Wu T. Felbamate as an add-on therapy for refractory epilepsy (Review). Cochrane Database Systematic Rev. 2011; 1; 1–13.
- Ryan K., Backos D. S., Reigan P. and Patel, M. Post-translational oxidative modification and inactivation of mitochondrial complex I in epileptogenesis. J Neurosci.2012; 32(33): 11250-11258.

- 31. Morimoto K., Fahnestock M. and Racine R.J. Kindling and status epilepticus models of epilepsy: rewiring the brain. Progr Neurobiol.2004; 73; 1–60.
- 32. Speranza L., Pesce M., Patruno A., Franceschelli S., de Lutiis M.A., Grilli A. and Felaco M. Astaxanthin treatment reduced oxidative induced pro-inflammatory cytokines secretion in U937: SHP-1 as a novel biological target. Mar Drugs.2012; 10(4); 890-9
- 33. Himmerich H., Bartsch S., Hamer H., Mergl R., Schönherr J., Petersein C. et al. Modulation of cytokine production by drugs with antiepileptic or mood stabilizer properties in anti-CD3- and anti-d40-stimulated blood in vitro. Oxid Med Cell Longev. **2014**; volume 2014: article ID 806162.
- 34. Yang W., Li J., Shang Y., Zhao L., Wang M., Shi J. and Li S. HMGB1-TLR4 Axis Plays a Regulatory Role in the Pathogenesis of Mesial Temporal Lobe Epilepsy in Immature Rat Model and Children via the p38MAPK Signaling Pathway. Neurochemical Research.2017; 42(4); 1179-1190.
- 35. Vezzani A., French J., Bartfai T. and Baram T.Z. (2011a): The role of inflammation in epilepsy. Nat Rev Neurol. 2011a; 7(1); 31-40.
- 36. Vezzani A., Maroso M., Balosso S., Sanchez M.A. and Bartfai T. IL-1 receptor/Toll-like receptor signaling in infection, inflammation, stress and neurodegeneration couples hyperexcitability and seizures. Brain Behav. Immun.2011b; 25 (7); 1281–1289.
- 37. Miller V. J., Karic V. V., Ofec R., Nehete S. R. and Smidt A. The temporomandibular opening index, report of headache and TMD, and implications for screening in general practice: an initial study. Quintessence Int.2014; 45(7); 605-612.
- 38. **Ravizza T., Gagliardi B., Noe F., Boer K., Aronica E. and Vezzani A.** Innate and adaptive immunity during epileptogenesis and spontaneous seizures: Evidence from experimental models and human temporal lobe epilepsy. Neurobiol Dis.**2008**; 29; 142–160
- Heinemann U., Kaufer D. and Friedman A. Blood-brain barrier dysfunction, TGFβ signaling, and astrocyte dysfunction in epilepsy. Glia. 2012; 60(8); 1251-1257.
- Xiao Z., Peng J., Yang L., Kong H. and Yin F. Interleukin-1β plays a role in the pathogenesis of mesial temporal lobe epilepsy through the PI3K/Akt/mTOR signaling pathway in hippocampal neurons. Journal of Neuroimmunology.2015; 282; 110-117
- 41. Vezzani A., Friedman A. and Dingledine R. J.

December. 2018 Vol. 24; Supplement Issue 2

The role of inflammation in epileptogenesis. Neuropharmacology.**2013**; 69; 16–24

- 42. Roseti C., Fucile S., Lauro C., Martinello K., Bertollini C., Esposito V., et al. Fractalkine/CX3CL1 modulates GABAA currents in human temporal lobe epilepsy. Epilepsia.2013; 54(10); 1834-1844.
- 43. Vezzani A. and Viviani B. Neuromodulatory properties of inflammatory cytokines and their impact on neuronal excitability. Neuropharmacology. 2015; 96(Pt A); 70-82.
- 44. Vezzani A. and Granata T. Brain inflammation in epilepsy: experimental and clinical evidence. Epilepsia.2005; 46(11); 1724-43.
- 45. Khadrawy Y.A., Sawie H.G. and Hosny E.N. Neuroprotective effect of curcumin nanoparticles against rat model of status epilepticus induced by pilocarpine. Journal of Complementary and Integrative Medicine.2018;15(3); 1-10.
- 46. **fujikawa D.G.** Activation of Caspase-Independent Programmed Pathways in Seizure-Induced Neuronal Necrosis. Fujikawa D. Acute Neuronal Injury. Springer, Cham Acute Neuronal Injury. **2018**; 11; 191-211.
- 47. **Teocchi M.A. and D'Souza-Li L.** Apoptosis through Death Receptors in Temporal Lobe Epilepsy-Associated Hippocampal Sclerosis. Mediators of Inflammation.**2016**; volume 2016. Article ID 8290562, 12 pages.
- 48. **Fujikawa D.G.** Prolonged seizures and cellular injury: understanding the connection. Epilepsy and Behavior.**2005**; 7(3); 3–11.
- 49. Hsu H. C., Tang N. Y., Liu C. H. and Hsieh C. L. Antiepileptic Effect of Uncaria rhynchophylla and Rhynchophylline Involved in the Initiation of c-Jun N-Terminal Kinase Phosphorylation of MAPK Signal Pathways in Acute Seizures of Kainic Acid-Treated Rats. Evid Based Complement Alternat Med.**2013**; Volume 2013. Article ID 961289.
- 50. **Henshall D. C.** Apoptosis signalling pathways in seizure-induced neuronal death and epilepsy. Biochem Soc Trans.**2007**; 35(2); 421-423.
- 51. Rajeshwari T., Raja B., Manivannan J., Silambarasan T. and Dhanalakshmi T. Valproic acid prevents the deregulation of lipid metabolism and renal renin-angiotensin system in L-NAME induced nitric oxide deficient hypertensive rats. Environ Toxicol Pharmacol. 2014; 37(3); 936-45
- 52. Cardinale J.P., Sriramula S., Pariaut R., Guggilam A., Mariappan N., Elks C.M., et al HDAC inhibition attenuates inflammatory, hypertrophic, and hypertensive responses in spontaneously hypertensive rats. Hypertension.

**2010**; 56; 437- 444.

- 53. Gómez C.D., Buijs R.M. and Sitges M.The anti-seizure drugs vinpocetine and carbamazepine, but not valproic acid, reduce inflammatory IL-1 $\beta$  and TNF- $\alpha$  expression in rat hippocampus. J Neurochem.**2014**; 130(6); 770-9.
- Cho G.W., Kang B.Y., Kim K.S. and Kim S.H. Effects of valproic acid on the expression of trophic factors in human bone marrow mesenchymal stromal cells. Neurosci Lett.2010; 526; 100–105.
- 55. Jung K.H., Han D.M., Jeong S.G., Choi M.R., Chai Y.G. and Cho G.W. Proteomic analysis reveals KRIT1 as a modulator for the antioxidant effects of valproic acid in human bone-marrow mesenchymal stromal cells. Drug Chem Toxicol.2015; 38(3); 286-92.
- 56. Robert V., Triffaux E., Savignac M. and Pelletier L. Singularities of calcium signaling in effector T-lymphocytes. Biochim Biophys Acta.2013; 1833(7); 1595-602.
- 57. Pérez M., Rojo A. I., Wandosell F., Díaz-Nido J. and Avila J. Prion peptide induces neuronal cell death through a pathway involving glycogen synthase kinase 3. Biochem J.2003; 372; 129–136
- 58. Chen Q., Ouyang D.y., Geng M., Xu L.h., Zhang Y.t., Wang F.p. and He X.h. Valproic acid exhibits biphasic effects on apoptotic cell death of activated lymphocytes through differential modulation of multiple signaling pathways. Journal of Immunotoxicology.2011; 8(3); 210-218.
- 59. Xie N., Wang C., Lin Y., Li H., Chen L., Zhang T., Sun Y., Zhang Y., Yin D. and Chi Z. The role of p38 MAPK in valproic acid induced microglia apoptosis. Neurosci Lett.2010; 482; 51–56
- 60. **Reddy D. S. and Jian K.** The testosteronederived neurosteroid androstanediol is a positive allosteric modulator of GABAA receptors. J Pharmacol Exp Ther. **2010**; 334(3); 1031-1041.
- Carver, C. M. and Reddy, D. S. Neurosteroid Structure-Activity Relationships for Functional Activation of Extrasynaptic δGABAA Receptors. The Journal of Pharmacology and Experimental Therapeutics. 2016; 357(1); 188– 204
- 62. Clossen B. L. andv Reddy D. S. Novel therapeutic approaches for disease-modification of epileptogenesis for curing epilepsy. Biochimica et Biophysica Acta. 2017; 1863(6); 1519–1538
- 63. Hosie A. M., Clarke L., da Silva H. and Smart T. G. Conserved site for neurosteroid

December. 2018 Vol. 24; Supplement Issue 2

modulation of GABA A receptors. Neuropharmacology.**2009**; 56(1); 149-154.

- 64. **Bromfield E.B., Cavazos J.E. and Sirven J.I.** An Introduction to Epilepsy. West Hartford (CT): Bromfield E.B., Cavazos J.E. and Sirven J.I. American Epilepsy Society. Clinical Epilepsy. Chapter 2.2006.
- 65. **Reddy D. S. and Rogawski M. A.** Ganaxolone suppression of behavioral and electrographic seizures in the mouse amygdala kindling model.Epilepsy Res.**2010**; 89(2-3); 254-260.
- 66. Zelcer N., Khanlou N. R., Jiang C.Q., Reed-Geaghan E.G., Landreth G.E., Vinters H.V and Tontonoz P. Attenuation of neuroinflammation and Alzheimer's disease pathology by liver x receptors. Proceedings of the National Academy of Sciences.2007; 104(25); 10601-10606
- 67. Saijo K.,Collier J.G., Li A.C., Katzenellenbogen J.A. and Glass C.K. An ADIOL-ERb-CtBP Transrepression Pathway Negatively RegulatesMicroglia-Mediated Inflammation Cell.2011; 145; 584–595.
- 68. **Hess J., Angel P. and Schorpp-Kistner M.** AP-1 subunits: quarrel and harmony among siblings". Journal of Cell Science. **2004**; 117 (Pt 25): 5965–73.
- 69. Groticke I., Hoffmann K. and Loscher W. Behavioral alterations in a mouse model of temporal lobe epilepsy induced by intrahippocampal injection of kainate. Exp Neurol. **2008**; 213; 71-83.
- 70. Dvkstra C.M. The development of neurodegeneration and behavioural alterations lithium/pilocarpine-induced after status epilepticus in rats. Institute of Medical Science of Toronto. University dysfunction and ultrastructural damage in the hippocampus.2012.
- 71. Covolan L. and Mello L.E. Assessment of the progressive nature of cell damage in the pilocarpine model of epilepsy. Braz J Med Biol Res.2006; 39; 915-924.
- 72. Moreira C.M., Masson S., Carvalho M.C. and Brandao M.L. Exploratory behaviour of rats in the elevated plus-maze is differentially sensitive to inactivation of the basolateral and central amygdaloid nuclei. Brain Res Bull.2007; 71; 466-473.
- 73. Carvalho M.C., Masson S., Brandao M.L. and de Souza Silva M.A. Anxiolytic-like effects of substance P administration into the dorsal, but not ventral, hippocampus and its influence on serotonin. Peptides. 2008; 29; 1191-1299
- 74. Zádori D., Geisz A., Vámos E., Vécsei L. and Klivényi P. Valproate ameliorates the survival

and the motor performance in a transgenic mouse model of Huntington's disease. Pharmacology Biochemistry and Behavior.**2009**; 94(1); 148-153.

- 75. **Tsutsui K. and Haraguchi S.** Breakthrough in neuroendocrinology by discovering novel neuropeptides and neurosteroids: 2. Discovery of neurosteroids and pineal neurosteroids. Gen Comp Endocrinol. **2014**; 205; 11-22.
- 76. **Borrow A. P. and Handa R. J.** Chapter Two -Estrogen Receptors Modulation of Anxiety-Like Behavior. Vitamins and Hormones. G. Litwack, Academic Press.**2017**; 103; 27-52.
- 77. Walf A. A and Frye C. A. Parity and estrogenadministration alter affective behavior of ovariectomized rats. Physiology & Behavior.2008; 93(1); 351–356.
- 78. Oyola M. G., PortilloW., Reyna A., Foradori C.D., Kudwa A., Hinds L., Handa R.J. and Mani S.K. Anxiolytic effects and neuroanatomical targets of estrogen receptorbeta (ER beta) activation by a selective ER beta agonist in female mice. Endocrinology.2012 153(2); 837-846.
- Nomura M., Akama K.T., Alves S.E., Korach K.S., Gustafsson J.A., Pfaff D.W. and Ogawa S. Differential distribution of estrogen receptor (ER)-alpha and ER-beta in the midbrain raphe nuclei and periaqueductal gray in male mouse: Predominant role of ER-beta in midbrain serotonergic systems. Neuroscience.2005; 130; 445–456.
- 80. Wu C.L., Huang L.T., Liou C.W., Wang T.J., Tung Y.R., Hsu H.Y. and Lai M.C. Lithium/pilocarpine-induced status epilepticus in immature rats result in long-term deficits in spatial learning and hippocampal cell loss. Neurosci Lett.2001; 312; 113-117.
- 81. Weihl C. C., Connolly A. M. and Pestronk A. Valproate may improve strength and function in patients with type III/IV spinal muscle atrophy.Neurology.2006; 67(3); 500–501.
- 82. Yu F., Wang Z., Tanaka M., Chiu C. T., Leeds P., Zhang Y. and Chuang D. M. Posttrauma cotreatment with lithium and valproate: reduction of lesion volume, attenuation of blood-brain barrier disruption, and improvement in motor coordination in mice with traumatic brain injury. J Neurosurg. (2013); 119(3); 766-773.
- 83. Esteves S., Duarte-Silva S., Naia L., Neves-Carvalho A., Teixeira-Castro A., Rego A.C., et al. Limited Effect of Chronic Valproic Acid Treatment in a Mouse Model of Machado-Joseph Disease. 2015; PLoS ONE 10(10): e0141610.

December. 2018 Vol. 24; Supplement Issue 2

- 84. Chou A. H., Chen S. Y., Yeh T. H., Weng Y. H. and Wang H. L. HDAC inhibitor sodium butyrate reverses transcriptional downregulation and ameliorates ataxic symptoms in a transgenic mouse model of SCA3. Neurobiol Dis.2011; 41(2); 481-488.
- Fan X., Xu H., Warner M. and Gustafsson, J. A. ERbeta in CNS: new roles in development and function. Prog Brain Res., 2012; 181; 233-250.
- Zeng-Li C., Guang-Li F., Chun-Mei Z. and Yi-Dan H. Effects of estrogen on ER, NGF and CHAT expression in cerebellum of aging female Sprague Dawley rat. Agric Sci China.2007; 6; 368–374.
- 87. Chakraborty T. R. and Gore A. C. Agingrelated changes in ovarian hormones, their receptors, and neuroendocrine function. Exp Biol Med (Maywood).2004; 229(10); 977-987.
- 88. **Hoppe C., Elger C. E. and Helmstaedter C.** Long-term memory impairment in patients with focal epilepsy. Epilepsia. **2007**; 48 (9); 26-29.
- 89. Groticke I., Hoffmann K. and Loscher W. Behavioral alterations in a mouse model of temporal lobe epilepsy induced by intrahippocampal injection of kainate. Exp Neurol.2008; 213; 71-83.
- 90. Chin J. and Scharfman H. E. Shared cognitive and behavioral impairments in epilepsy and Alzheimer's disease and potential underlying mechanisms. Epilepsy & Behavior : **2013**; E&B. 26(3); 343–351.
- 91. **Park S.P. and Kwon S.H.** Cognitive Effects of Antiepileptic Drugs. Journal of Clinical Neurology (Seoul, Korea).**2008**; 4(3); 99–106.
- 92. **Sgobio C., Ghiglieri V., Costa C. et al** Hippocampal synaptic plasticity, memory, and epilepsy: effects of long-term valproic acid treatment. Biol Psychiat.**2010**; 67; 567–574.
- 93. Aboul Ezz H. S., Khadrawy Y. A and Noor, N. A. The Neuroprotective Effect of Curcumin and Nigella sativa Oil Against Oxidative Stress in the Pilocarpine Model of Epilepsy: A Comparison with Valproate. Neurochemical Research.2011; 36(11); 2195-2204.
- 94. Bodo C. and Rissman, E. F. New roles for estrogen receptor  $\beta$  in behavior and neuroendocrinology. Frontiers in Neuroendocrinology. **2006**; 27(2); 217-232.
- 95. Hampson E. and Kimura D. Reciprocal effects of hormonal fluctuations on human motor and perceptual-spatial skills. Behav Neurosci.**1998**; 102(3); 456-459.
- 96. **Phillips S. M. and Sherwin, B. B.** Variations in memory function and sex steroid hormones across the menstrual cycle.

December. 2018 Vol. 24; Supplement Issue 2

Psychoneuroendocrinology.**1992**; 17(5); 497-506

97. Maki P. M., Gast M. J., Vieweg A. J, Burriss S. W. and Yaffe K. Hormone therapy in menopausal women with cognitive complaints: a randomized, double-blind trial. Neurology.2007; 69(13); 1322-1330.