

 https://doi.org/10.21608/zumj.2023.207221.2792

 Manuscript ID
 ZUMJ-2304-2792 (R1)

 DOI
 10.21608/ZUMJ.2023.207221.2792

Volume 29, Issue 5, - September 2023

ORIGINAL ARTICLE

The Role of Serum Glutathione Peroxidase Enzyme in Relapsing-Remitting Multiple Sclerosis.

Bothina M. Ramadan¹, Yosria A Altaweel¹, Khaled Aly El Sharkawyv¹, Nada Reda Elgamasy² 1 Neurology Department, Faculty of Medicine, Zagazig University, Zagazig, Egypt

2 Neurology Department, Al-Ahrar Teaching Hospital; Zagazig, Sharkia, Egypt

Corresponding author Bothina M Ramadan E-mail: drbothinaramadan@gmail.com

 Submit Date
 2023-05-31

 Revise Date
 2023-06-15

 Accept Date
 2023-06-25

ABSTRACT

Background: Multiple sclerosis (MS) is considered an immunemediated disorder with different disease course., Recently, oxidative stress was reported to play a role in the pathogenesis of MS. Aim: to investigate the role of the serum glutathione peroxidase enzyme in the pathogenesis of Multiple sclerosis, especially the relapsing-remitting type (RRMS), and to assess the relationship between it and disease characteristics.

Methods: 45 patients with RRMS (23 in relapse and 22 in remission) (18 males and 27 females) and 45 age, sex, and body mass index (BMI)-matched healthy subjects were included in this study. Clinical assessment, general, and neurological examination were done. MS Severity was calculated using the expanded disability status scale (EDSS). Magnetic resonance imaging (MRI) of the brain and spine with gadolinium enhancement and visual evoked potential was done for all MS patients. Serum glutathione peroxidase (GP_X) enzyme and routine laboratory tests were assessed.

Results: reduced levels of GP in patients compared to controls. No significant difference was detected between the remitting and relapsing MS groups. In addition, there was a statistically significant negative correlation between serum GPx, disease duration, and physical MSIS. Also, an adverse correlation between serum GPx and

gadolinium enhancement was recorded.

Conclusion: Our study demonstrates a significantly lower level of GPX among patients than among controls; this indicates its role in MS pathogenesis, which might be



important for new therapeutic strategies based on an antioxidant approach in MS patients.

Keywords: Relapsing-Remitting Multiple Sclerosis, glutathione peroxidase, pathogenesis.

INTRODUCTION

Multiple sclerosis (MS) is characterized by inflammation, demyelination, and axonal loss in the central nervous system (CNS) (1). It affects around 2.5 million individuals worldwide. Oxidative stress results from an imbalance between oxidant and antioxidant systems and is characterized by excess free radical production e.g. reactive oxygen species (ROS) and reactive nitrogen species (RNS) causing injury or cell death (2). These antioxidant molecules include vitamin E, vitamin D, glutathione, antioxidant enzymes (superoxide dismutase, catalase, glutathione), and melatonin (2). In MS, ROS is also considered a marker of oxidative stress as evidenced by the elevation of 7-ketocholesterol in the CSF of MS patients (3). Also, Nitric oxide (NO) and its metabolites were reported to be elevated in the blood and CSF of MS patients. (4)Glutathione peroxidase (GPX), one of the peroxidase family, is considered one of the endogenous antioxidants that protect against oxidative damage (5). The main role of GPX is to reduce both free oxygen peroxide and lipid hydroperoxides to H2O(5), the enzyme has 8 isoenzymes: GPX1 is the highly abundant form, GPX 2 is found to be of lower levels in relapsing-remitting MS (RRMS) as lower glutathione levels point to the presence of oxidative stress in MS (6). This research aims to examine the role of serum glutathione peroxidase enzyme in the pathogenesis of RRMS and to explore the relation between it and disease characteristics.

METHODS

Ethics approval and consent to participate: The study was approved by the Ethical Committee of our Faculty (ZU-IRB# 4164/22-11-2017). Written informed consent was obtained from all patients who agreed to participate in our study.

This case-control study was conducted in the neurology department and MS outpatient clinic at Zagazig University Hospitals in the period from February 2019 to October 2021. The study included 90 subjects divided into 45 patients (18 males and 27 females) and 45 age, sex, and BMImatched nonsmoker healthy controls without any neurological diseases. The study was done according to The Code of Ethics of the World Medical Association (Declaration of Helsinki) for studies involving humans. The study's Ethical approval number was (ZU-IRB# 4164/22-11-2017). Written consent was obtained from all participants in our study. RRMS was diagnosed according to the recently modified McDonald criteria 2017 (7).

Patients with the relapsing-remitting disease were classified into 2 groups as follows: Group (A): (Remission group): If patients presented 1 month after the relapse and when symptoms had become stable (8) and included 22 patients. Group (B): (Relapse group): If there were new symptoms or there was a reoccurrence of old symptoms, for at least 24 hours and occurred after 30 days or more from the previous relapse in the absence of an alteration in core body temperature or infection and included 23 patients. Expanded disability status score (EDSS) is between 1.5- 6 to include ambulatory patients (9). No antioxidant drugs were used by the patients.

The control group included 45 subjects in our study of age, sex, and BMI-matched nonsmoker healthy persons. We excluded patients with a history of other neurologic illnesses i.e. strokes, brain tumors, neurodegenerative disorders, and other demyelinating diseases. Other concomitant diseases such as diabetes mellitus, hypertension, hepatic, renal, and pulmonary diseases as well as patients with present infection and pregnant females were excluded also.

Patients were subjected to full medical and neurological history, complete general and neurological examination, criteria of MS (duration of disease, annualized relapse rate, severity of relapses, type of treatment used during relapse and remission) were assessed, annualized relapse rate was calculated by dividing number of relapses by duration of disease (**10**). We got annualized relapse rate by dividing the number of relapses by the duration of the disease (10). Assessment of disease disability was assessed using EDSS (9). Evidence of disease activity was assessed using the Evidence of disease activity scale and criteria which include physical and psychological Multiple Sclerosis impact scale (11).

Radiological investigations including magnetic resonance imaging (MRI) of the brain and spinal cord were conducted. The MS imaging protocol was performed using a 1.5 tesla superconducting MR imager (Achieva, Philips Medical System).

Complete blood count (CBC), hepatic and renal function, lipid profile, ESR, CRP, random blood sugar, and serum uric acid were done as a routine investigation. Special laboratory investigation: enzyme-linked immunosorbent assay (ELISA) for glutathione peroxidase detection using Tecan Austria GmbH 5082 Grodig, Austria SUNRISE (2002).

STATISTICAL ANALYSIS

Data were tabulated and SPSS was used for analysis (12). Data were presented as mean \pm standard deviation (SD), frequencies, and percentages. Student t-test, ANOVA, or chi-square test was performed when suitable. Correlation analysis was performed. P < 0.05 was considered statistically significant (S).

RESULTS

MS was higher in females than males in a ratio of 1.5:1 nearly, with the mean age (\pm SD) of the remission group was 37.7 (\pm 9.5) years and the relapse group was 34.2 (\pm 8.7) (table 1). The GPx level was reduced in patients than control but there is no difference in its level between remitting and relapsing groups (table 1).

MS duration (M±SD) was 2.1 ± 2.4 years and the mean ARR was $2.1 (\pm 1.9)$. 80% of cases had a total relapse rate (TRR) less than 5 while 20% of cases had TRR equal or more than 5. EDSS less than 5 in 82% of cases, and only 18% have EDSS equal or more than 5.

Regarding the initial manifestations, the majority of cases (44.4%) presented with pyramidal manifestations followed by visual manifestations (24%). 22.2% of cases presented with sensory manifestations. Brain stem or cerebellar manifestations were initial in 4.4% of cases (table 2).

Table (3) shows that serum GPx levels didn't differ significantly between males and females and also between patients in remission and relapse. Moreover, there is no significant difference in its level as regards ARR or treatment. In addition, GPx was statistically significantly higher in patients treated with fingolimod and ocrelizumab (table 4).

As regard correlation, there was a negative correlation between serum GPx and disease duration, and physical MSIS. Also, there was an adverse correlation between serum GPx and gadolinium enhancement, but no significant **Table 1:** Demographic data of studied groups

correlation between GPX and VEP latency and amplitude and other laboratory markers (Table 5; Figures 1 and 2). No significant correlation between GPx and markers of inflammation (WBCS (total and differential), CRP) or other laboratory biomarkers could be observed (Table 6)

Demographic data	Rem N=22	ission 2	Rela N=2	apse 23	Cont N=4		Test of significance	Р
Age (Years)								
Mean±SD	37.7	±9.5	34.2	±8.7	33.9	±10.8	F=1.06	0.34
Range	17-50	17-50		17-47		5		
Sex	Ν	%	Ν	%	Ν	%	χ2=1.36	0.5
Male	7	31.8	11	47.8	20	44.4		
Female	15	68.2	12	52.2	25	55.6		
BMI (KG/m2)							-	
X (±SD)	29.2	±4.2	30.2	±3.8	29.0	±3.6	F=0.7	0.49
Range		5	20-3	35.5	19-3	9		
Glutathione peroxidase (ng/ ml)	100.5	5±18.7	102	4±11	120.0	5±29.3	F=7.47	0.001*

t: student's t-test, X2: chi-square test, X: mean, SD: Standard Deviation, BMI: Body Mass Index, F: Analysis Of Variance (ANOVA).

Table 2: Clinical Characteristics of MS Patients

	Range	Mean±SD
Duration of disease in years	0.25-8	2.1±2.4
ARR	0.5-8	2.1±1.9
Total relapse rate	Ν	%
<5	36	80
_≥5	9	20
Initial manifestations	Ν	%
Pyramidal manifestation	20	44.4
sensory manifestation	10	22.2
cerebellar manifestation	2	4.4
Brain stem manifestation	2	4.4
Visual manifestation	11	24
EDSS	Ν	%
<5	37	82
≥5	8	18
	$1 \mathbf{D}^{\prime} + 1^{\prime} 1^{\prime} + 0^{\prime} + \mathbf$	

ARR: Annualized relapse rate and EDSS: Expanded Disability Status Scale

Table 3: Difference between serum GPx levels in males and females, and its relation with Remission, Relapses, ARR, and treatment

Parameter	Ν	GPx (ng/ml)	t-test	Р
		$\overline{X}_{\pm SD}$		
Gender				
Male	18	106.1±12.4	1.68	0.09
Female	27	90.4 ± 16.5		
Remission	22	100.5±18.8	0.42	0.67
Relapse	23	102.5±11.5		
ARR	·		•	
>1	29	100.2 ± 14.6	0.78	0.43
<1	16	103.97 ± 16.7		
Treatment				
Non-specific	18	101.2±15.9	0.17	0.85
MS-specific therapy	27	102 ± 14.9		
: student t-test, X: mean, SD: Standard	Deviation, and ARR:	Annual Relapse F	Rate	•

Ramadan, B., et al

Table 4: Serum glutathione peroxidase in different types of disease-modifying drugs

DMD	Gpx (ng/ml)	F	Р
	$\overline{\mathrm{X}}_{\pm \mathbf{SD}}$		
Nonspecific	95.8±14.	3.5	0.01*
IFN B	104.6±13.1		
Fingolimod	109.34±0.0		
Ocrelizumab	109.3±8.3		
Cyclophosphamide	97.2±4.5		
Rituximab	53.1±0.0		

DMD: disease-modifying drugs, **X**: mean, **SD:** Standard Deviation, **F:** Analysis of Variance (ANOVA) * significant.**EDSS:** Expanded Disability Status Scale, **CRP:** C reactive protein and NS: non-significant

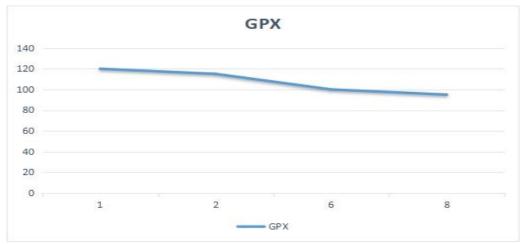
Table	5:	Correlation	of	serum	Glutathione	peroxidase	with	clinical,	radiological,	and
electrop	ohysio	ological marke	rs							

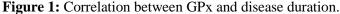
Parameter	R	Р
Disease duration (years)	-	< 0.05
•	0.34	*
ARR	-0.06	NS
EDSS	-0.25	NS
MSIS physical	-0.32	< 0.05*
MSIS psychological	-0.28	NS
Gadolinium enhancement	-0.33	< 0.05
		*
New T2 lesions	-0.23	NS
Cervical patches	0.21	NS
Black holes	-0.05	NS
Brain atrophy	-0.04	NS
VEP latency		
.Rt	0.009	NS
.Lt	0.006	NS
VEP Amplitude		
Rt.	0.09	NS
Rt.	0.01	NS

Table 6: Correlation between GPx and laboratory biomarkers

Parameter	R	Р
WBCs	0.07	NS
Neutrophils	0.08	NS
Monocytes	0.03	NS
Lymphocytes	0.02	NS
CRP	0.07	NS
Cholesterol(mg/dl)	0.1	NS
Triglyceride (mg/dl)	0.09	NS
LDL(mg/dl)	0.003	NS
HDL (mg/dl)	-0.08	NS
RBS	0.08	NS
Albumin	0.13	NS
ALT	0.07	NS
AST	0.001	NS
Uric acid	0.17	NS

WBCs: complete blood count; CRP: C-reactive protein; LDL: low-density lipoprotein; HDL: high-density lipoprotein; RBS: Random blood sugar; ALT: Alanine transaminase; AST: Aspartate transaminase





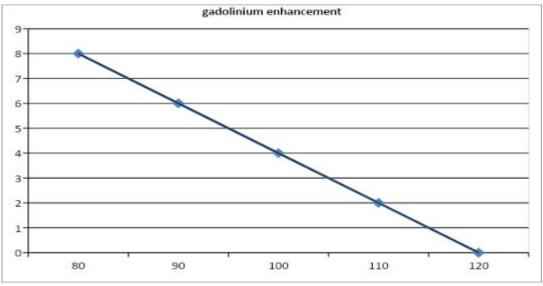


Figure 2: Correlation between GPx and gadolinium enhancement.

DISCUSSION

As a multi-factorial disease, multiple sclerosis (MS) characterized by an interaction between genetic predisposition, environmental factors, and aberrant immune response was reported (13, 14). The impact of Oxidative stress in MS pathogenesis has been reported previously as there were increased blood concentrations of the reactive oxygen species causing cell injury and death (15, 16, 17, 18). Glutathione (GSH), an endogenous antioxidant, is a vital agent in the detoxification of their xenobiotics and metabolites (19), detoxification of reactive oxidants (20), and preservation of the intracellular redox balance which is very important in the brain (21). It acts as a co-substrate to glutathione peroxidase (GPX) and glutathione reductase (GR) enzymes. (22). Glutathione peroxidase catalyzes the reduction of hydroperoxides such as H2O2 to oxidized glutathione (GSSG) to protect cells against oxidative damage and GSSG can also be converted back to GSH by glutathione reductase (GR) (2). Some studies verified normal antioxidant activity

in blood, while others indicate a decrease or increase in this activity.

In our study, we found a significantly decreased serum level of glutathione peroxidase in patients than in the control group. This finding is consistent with the result of earlier studies (5, 6, 13, 23, 24) that reported GR and GSH concentrations were higher in sera of MS patients than controls is consequently associated with a decrease in GPX. Elevation of GSH was verified by stimulation of extracellular glutamate which represents that GSH has a neuroprotective role against glutamate toxicity and oxidative damage. Tasset and colleagues found higher activity of GR in RRMS patients compared to controls. In addition, an increased ratio of reduced glutathione to oxidized glutathione (GSH/GSSG) in these patients was recorded (5). On the contrary, the results obtained by Calabrese and colleagues; Krotenko and colleagues Ortiz and colleagues, and Choi and colleagues reported decreased GSH and increased GPX (25, 26). However antioxidants levels increase approving of the theory that in an attempt to protect cells, the cellular response to oxidative stress is likely to arise before the inflammatory response, and when this response is inadequate then a relapse in MS happens (5). Ortiz and colleagues showed that GSH level is lowered by 40–50% in RRMS patients than controls, thus GPx levels increased (27). However, Di Giussepe and colleagues demonstrated no change in GSH among MS patients (28).

Oxidative Stress gained much importance in MS throughout the disease course. Inflammatory processes are initiated by it during the acute phase, while neurodegeneration is maintained by it in the chronic phase (1). In this study, we noticed a negative correlation between GPX and disease duration that goes in line with Acar and colleagues (29); Tasset and colleagues (5), and Ferriera and colleagues (2). Fischer and colleagues reported that increased oxidative stress damage leads to more disease severity (30), EDSS is the most widely used index of MS severity. In this study, we found no statistical significance correlation between GPX and EDSS. In contrast, Acar and colleagues (29), Tasset and colleagues (5), and Girona and colleagues (18) found depletion of GSH, GSH/GSSG ratio, and a decline in GSSG levels and increased GPX in benign MS. This finding could suggest that antioxidant diminution is a probably causative factor not a consequence of MS severity. Miller and colleagues (**31**) and Ljubisavljevic and colleagues (32) found a negative correlation between oxidative stress markers and EDSS. On the other hand, we found a significant negative correlation between GPX level and physical MSIS, this coincides with Adamczyk and Adamczyk-Sowa that reported a decrease in MSIS after oxidative stress repair (33).

Regarding the relation between GPX and MRI imaging, we found an inverse correlation between GPX and gadolinium enhancement lesions. This matches with Miller and colleagues (34) and Adamczyk and Adamczyk-Sowa's results (33). In contrast, Ljubisavljevic and colleagues did not report such a correlation (31).

The effect of MS pharmacotherapy on biomarkers of oxidative stress has been studied previously, fingolimod and ocrelizumab were the highly studied medications. We found a positive correlation between GPX and treatment with fingolimod and ocrelizumab. Similarly, Yevgi and Demir found that fingolimod decreases total oxidative stress (TOS) and also decreases pre and post-treatment EDSS (**35**). In contrast, Pegoretti and colleagues found that ocrelizumab does not affect oxidative stress (**36**). Moreover, previous studies found no correlation between oxidative stress and types of drug treatment (18, 29).

In this study, although we observed a significant difference in WBCs between relapse and remission groups, we found a negative correlation between GPX and signs of inflammation but with no statistical significance. Similar to our results, Koch and colleagues (37); Miller and colleagues (34); Witherick and colleagues (38); Castegna and colleagues (39), and Usatyuk and Natarajan (40) reported no significant correlation between oxidative stress and either disease severity or inflammatory activity. This may be explained as oxidative stress heralds the response, thus backing inflammatory the proposition that oxidative stress could modify the permeability of the blood-brain barrier and enhances the adhesion of monocytes to the vascular endothelium.

CONCLUSION

GPX is higher in patients with RRMS than control, but no statistically significant difference was observed in remission or relapse patients, this indicates its role in pathophysiology. We recommend its use as a new therapeutic strategy in the protection and treatment of MS.

REFERENCES

- Adamczyk-Sowa M, Sabina G, Ewa Ż, Michalina G, Katarzyna N, Paweł S, et al. The influence of sodium on pathophysiology of multiple sclerosis. J Neurol Sci. 2017; 38: 389–398.
- Ferreira B, Mendes F, Osorio N, Caseiro A, Gabriel A and Valado A. Glutathione in multiple sclerosis. J Biomed Sci. 2013; 70 (2): 75-9.
- 3. Diestel A, Aktas O, Hackel D, Hake I, Meier S, Raine CS, et al. Activation of microglial poly (ADPribose)-polymerase-1 by cholesterol breakdown products during neuroinflammation: a link between demyelination and neuronal damage. J Exp Med. 2003; 198 (11): 1729–40.
- Rejdak K, Eikelenboom MJ, Petzold A, Thompson EJ, Stelmasiak Z, Lazeron RH, et al . CSF nitric oxide metabolites are associated with activity and progression of multiple sclerosis. Neurol. 2004; 63 (8): 1439–45.
- 5. Tassets I, Aguera E, Sanchez-Lopez F, Feijoo M, Giraldo AI, Cruz AH, et al. Peripheral oxidative stress in relapsing-remitting multiple sclerosis. J Clin Biochem. 2012; 45: 440–44.
- Socha K, Kochanowicz J, Karpińska E, Soroczyńska J, Jakoniuk M, Mariak Z, et al. Dietary habits and selenium, glutathione peroxidase and total antioxidant status in the serum of patients with relapsing-remitting multiple sclerosis. Nutr J. 2014; 13: 62–6.
- Thompson AJ, Banwell B L, Barkhof F, Carroll WM, Coetzee T, Comi G, et al. Diagnosis of multiple sclerosis: 2017 revisions of the McDonald criteria. The Lancet Neurol. 2018; 17 (2): 162-73.

- 8. Poser CM, Paty DW, Scheinberg L, McDonald WI, Davis FA, Ebers GC, et al . New diagnostic criteria for multiple sclerosis: Guidelines for research protocols. Ann Neurol. 1983; 13: 227–31.
- Cao H, <u>Peyrodie</u> L, BoudetS, Cavillon F, Agnani O, Hautecoeur P, et al. Expanded Disability Status Scale (EDSS) estimation in multiple sclerosis from posturographic data. Gait & Posture. 2013; 37 (2) 242–45.
- 10. Steinvorth SM, Röver C, Schneider S, Nicholas R, Straube S and Friede T. Explaining temporal trends in annualized relapse rates in placebo groups of randomized controlled trials in relapsing multiple sclerosis: systematic review and meta-regression. Mult Scler J. 2013; 17: 1211-17.
- 11. Winder GL and Allen DD .Measurement characteristics and clinical utility of 29–item Multiple Sclerosis Impact Scale. Arch Phys Med Rehabil. 2014; 95: 593-94.
- 12. IBM Corporation: IBM SPSS statistics for Windows version 24.0 Armnok, NY: IBM Corp; 2016.
- 13. Zhang C, Zhang M and QiuW .Safety and efficacy of tocilizumab versus azathioprine in highly relapsing neuromyelitis optica spectrum disorder (TANGO): an open-label, multicentre, randomised, phase 2 trial. Lancet Neurol. 2020; 19: 391–401.
- 14. Chastain EM and Miller SD .Molecular mimicry as an inducing trigger for CNS autoimmune demyelinating disease. Immunol Rev. 2012; 245 (1): 227-38.
- 15. Van-Horssen J, Brink BP, De-Vries HE, van der Valk, P and Bo L. The blood-brain barrier in cortical multiple sclerosis lesions. J. Neuropathol. Exp. Neurol. 2011; 66: 321–28.
- 16. De Riccardis L, Buccolieri A, Muci M, Pitotti E, De Robertis F, Trianni G, et al . Copper and ceruloplasmin dyshomeostasis in serum and cerebrospinal fluid of multiple sclerosis subjects. Biochim. Biophys. Acta Mol. Basis Dis. 2018; 18 (5): 1828–38.
- 17. Herrmann JM and Dick TP. Redox biology on the rise. Biol.Chem. 2012; 393 (9): 999–1004.
- 18. Gironi M, Borgiani B, Mariani E, Cursano C, Mendozzi L, Cavarretta R, et al. Oxidative stress is differentially present in multiple sclerosis courses, early evident, and unrelated to treatment. J Immunol Res. 2014; 96 (1): 10–11.
- GuFeng, ChauhanVedand Chauhan and Abha . Glutathione redox imbalance in brain disorders. Curr Opin Clin Nutr Metab Care. 2015; 18 (1): 89-95
- 20. Carvalho AL, Lim JL, Nijland P, Witte ME and Horssen JV. Glutathione in multiple sclerosis: More than just an antioxidant? Mult Scler J. 2014; 23 (2): 12-24.
- 21. Aoyama K and Nakaki T .Neuroprotective properties of the excitatory amino acid carrier 1 (EAAC1) Amino Acids. Springer. 2013; 45 (1): 133–42.
- 22. Tavassolifar MJ, Vodjgani M, Salehi Z and IzadM .The Influence of Reactive Oxygen Species in the Immune System and Pathogenesis of Multiple Sclerosis. Autoimmune Dis. 2020; 6 (2): 12-20.
- Ramadan, B., et al

- Miller E, Mrowicka M, Zolynski K and KedzioraJ .Oxidative stress in multiple sclerosis (in Polish). Pol Merkur Lekarski. 2009; 27 (162): 499–502.
- 24. Srinivasan R, Ratiney H, Hammond-Rosenbluth KE, Pelletier D and Nelson SJ. MR spectroscopic imaging of glutathione in the white and gray matter at 7 T with an application to multiple sclerosis. Magn. Reson Imaging. 2010; 28: 163–70.
- 25. Ortiz GG, Macı'as-Islas MA, Pacheco-Moise's FP, Cruz-Ramos JA, Sustersik S, BarbaEA, et al . Oxidative stress is increased in serum from Mexican patients with relapsing remitting multiple sclerosis. Dis Markers 2009; 26: 35–9.
- 26. Choi IY, Lee SP, Denney DR and Lynch SG. Lower levels of glutathione in the brains of secondary progressive multiple sclerosis patients measured by 1H magnetic resonance chemical shift imaging at 3 T. Mult Scler J. 2011; 17: 289–96.
- 27. Ortiz GG, Pacheco-Moisés FP, Torres-Sánchez ED, Sorto-Gómez TE, Mireles-Ramírez M, León-Gil A, et al . Multiple Sclerosis and Its Relationship with Oxidative Stress, Glutathione Redox System, ATPase System, and Membrane Fluidity. Trending topics in Multiple Sclerosis. 2016; (3): 44-9.
- 28. Di Giuseppe D, Ulivelli M, Bartalini S Battistini S, Cerase A, PasseroS, et al . Regulation of redox forms of plasma thiols by albumin in multiple sclerosis after fasting and methionine loading test. Amino Acids. 2010; 38 (5): 1461–71
- 29. Acar A, CevikMU, Evliyaoglu O, Uzar E, Tamam Y, Arıkanoglu A et al .Evaluation of serum oxidant/antioxidant balance in multiple sclerosis. Acta Neurol Belg. 2012; 3: 275–80.
- 30. Fisher E, Rudick RA, Simon JH, Cutter G, Baier M, Lee JC, et al . Eight-year follow-up study of brain atrophy in patients with MS. Neurol. 2002; 59 (9): 1412–20.
- 31. Miller E, Walczak A, Majsterek I and KedzioraJ .Melatonin reduces oxidative stress in the erythrocytes of multiple sclerosis patients with secondary progressive clinical course. J Neuroimmunol. 2013; 257: 97–101.
- 32. <u>Ljubisavljevic</u> S, <u>StojanovicI,Tatjana</u> C, <u>Vojinovic</u> S, Stojanov D, Stojanovic D, et al .Erythrocytes' antioxidative capacity as a potential marker of oxidative stress intensity in neuroinflammation. J Neurol Sci. 2014; 337 (2): 6-11.
- 33. Adamczyk B., Adamczyk-Sowa M. New insights into the role of oxidative stress mechanisms in the pathophysiology and treatment of multiple sclerosis. Oxid Med Cell Longev. 2016; 20 (2): 95–8.
- 34. Miller E, Walczak A, Saluk, J, Ponczek M B and Majsterek I. Oxidative modification of patient's plasma proteins and its role in pathogenesis of multiple sclerosis," Clin Biochem. 2012; 45 (2): 26– 30.
- 35. Yevgi R and DemirR .Oxidative stress activity of fingolimod in multiple sclerosis. J Clin Neurol Neurosurg. 2021; 20 (2): 106-200.
- 36. Pegoretti v, Swanson KA, Bethea JR, ProbertL, Eisel UM and FischeR . Inflammation and Oxidative Stress in Multiple Sclerosis: Consequences for

Therapy Development. Oxid Med Cell Longev. 2020; 7191080.

- 37. Koch M, Ramsaransing GS, Arutjunyan AV, Stepanov M, Teelken A, Heersema DJ, et al . Oxidative stress in serum and peripheral blood leukocytes in patients with different disease courses of multiple sclerosis. J Neurol. 2006; 253: 483–87.
- 38. Witherick J, Wilkins A, Scolding N and Kemp K. Mechanisms of oxidative damage in multiple sclerosis and a cell therapy approach to treatment. Autoimmune Dis. 2010; (2): 12-19.
- 39. Castegna A, Palmieri L, Spera I, Porcelli V, Palmieri F, Fabis-Pedrini MJ, et al . Oxidative stress and

To Cite

Ramadan, B., altaweel, Y., Elsharkawy, K., Elgamasy, N. THE ROLE OF SERUM GLUTATHIONE PEROXIDASE ENZYME IN RELAPSING-REMITTING MULTIPLE SCLEROSIS. Zagazig University Medical Journal, 2023; (1339-1346): -. doi: 10.21608/zumj.2023.207221.2792

- reduced glutamine synthetase activity in the absence of inflammation in the cortex of mice with experimental allergic encephalomyelitis. Neurosci. 2011; 185: 97–105.
- 40.Usatyuk PV and Natarajan V .Hydroxyalkenals and oxidized phospholipids modulation of endothelial cytoskeleton, focal adhesion and adherens junction proteins in regulating endothelial barrier function. Microvasc Res. 2012; 83 (1): 45–55.